

The Impact of the “Low Molecular Weight Heparin” Administration on the Clinical Course of the COVID-19 Disease

Background: Lymphopenia is the most important criterion of the mortality and discharging feature for patients infected with coronavirus disease 2019 (COVID-19). This study aimed to investigate the clinical impact of a low molecular weight heparin (LMWH) treatment on the clinical course of COVID-19.

Materials and Methods: Patients’ clinical symptoms, radiologic outcomes, hematologic, biochemical, D-dimer, and C-reactive protein (CRP) results were obtained from medical records. Participants were separated into two groups: one was treated with LMWH and the other was not. Improvement in the patients was compared before and after treatment.

Results: Ninety-six patients who were diagnosed with COVID-19 between April and May 2020 were retrospectively analyzed. The multivariable analysis showed that the count of lymphocytes, D-dimer, and CRP levels were significantly improved in the LMWH group, as compared to the control group (OR, (95% CI) 0.628 (0.248–0.965), $p < 0.001$), OR, (95% CI) 0.356 (0.089–0.674), $p < 0.001$, respectively). The area under the receiver operating characteristic (ROC) curve analysis was AUC: 0.679 ± 0.055 , 0.615 ± 0.058 , 0.633 ± 0.057 , respectively, the β value was found as -1.032, -0.026, and -0.465, respectively.

Conclusion: LMWH treatment group demonstrated better laboratory findings, including recovery in the lymphocyte count, CRP, and D-dimer results.

Keywords: Coronavirus disease-19 (COVID-19), heparin, anti-inflammatory, lymphocyte

Introduction

The COVID-19 disease is an illness caused by a respiratory and systemic zoonotic coronavirus. It was first recognized in Wuhan, China, and it has continued to spread rapidly since then. It has become a global calamity, and the World Health Organization (WHO) declared the disease a pandemic on March 11, 2020. Over two million people worldwide have been infected so far, and the mortality continues to rise. However, no effective medical cure is available, and patients are treated according to their symptoms and findings [1,2].

Helpful strategies for improvement from the illness may be devised if the pathophysiology is understood. Lymphopenia and cytokine storms are the typical pathological changes detected in patients with coronavirus infections; they relate to the COVID-19 severity. The cytokine storm is a key mechanism underlying the disease exacerbation and mortality in COVID-19 patients [3–5]. Some studies have indicated that LMWH has some properties other than its anticoagulant effects, such as its anti-inflammatory action, which ensures improvement in lymphopenia[6]. However, although the anti-inflammatory effects of LMWH in COVID-19 remain unclear, it is thought that the anti-inflammatory efficacy contributes to the regression of COVID-19 [7]. The aim of this study is to assess the clinical impact of LMWH treatment on the clinical course of COVID-19.

Material and Methods

In this study, 96 participants who were admitted to the Selcuk University Hospital, Department of Pulmonology, **between March and April 2020**, were analyzed retrospectively. The patients' clinical outcomes were investigated by means of electronic medical records. COVID-19 was diagnosed according to the WHO guidelines. This research was approved by the Ministry of Health committee (approval number: 2020/37) and conducted in accordance with the principles of the Declaration of Helsinki [2].

The blood samples were gathered during the length of the stay in the hospital: D-dimer, prothrombin time (PT), international normalized ratio (INR), and fibrinogen (FIB) measurements were investigated by utilizing a coagulation analyzer device.

Participants were separated into two groups according to D-dimer (D-dimer > 750 ng/mL) and PT (<12 sec) outcomes due to mortality, raised accordingly with D-dimer, PT levels, and applied appropriate thrombopropylaxis with LMWH (thromboprophylactic dose of 4000 UI/day, for 7 days). No anticoagulant drugs other than heparin were utilized for seven days or longer in the research patients. All of the participants received appropriate treatment after admission to the hospital [8–10].

Inclusion Criteria

The inclusion criteria included 1) satisfying the conditions of the diagnostic standards of COVID-19 pneumonia explained by the Health Commission of Turkey, 2) identified as tightness of breath, respiration rate (RR) \geq 30/minute, detected typical lesions in CT images of viral pneumonia, 3) age \geq 18 years, 4) no history of any pulmonary disease, and 5) no immunosuppressive or corticosteroid agent during the therapy period.

Exclusion Criteria

The exclusion criteria included 1) participants with severe chronic diseases, 2) patients who had liver, kidney, or cardiac disease, 3) patients who had taken LMWH therapy for the last three months, 4) patients who had a history of mental disorders, 5) women who were pregnancy or breastfeeding, 6) patients who had followed-up in the intensive care unit (ICU), and 7) patients with any sensitivity to LMWH.

Chest CT Severity Score Assessment

The computer tomography severity score (CT-SS) was utilized to evaluate the patients with COVID-19. The CT-SS is an adapted version of the scoring system describing ground glass, interstitial opacity, and air trapping for severe acute respiratory syndrome (SARS). This scoring system assesses pulmonary lesions as a guide for detecting the disease [11]. The lungs were separated into twenty zones, according to the anatomy of the human. The detected pulmonary opacities in all zones were assessed by thin section thorax CT (TST-CT) using system linked scores of 0, 1, and 2, according to opacification, including 0%, <50%, or \geq 50% of every zone. The total scores, which ranged between 0 and 40 points, were obtained from each zone that was collected. Two experienced thorax radiologists, who were blinded to the identity of the participants, evaluated all of the CT screenings. Chest CT scans were performed with a 256-detector CT scanner (Siemens, Germany). All of the participants were lying in the supine position, and the scan was performed during the breath hold situation [11].

Data Collection

The demographic characteristics (age, gender, BMI), signs and symptoms, clinical outcomes, initial knowledge, complete blood count (CBC), D-dimer, FIB, coagulation profile, inflammatory markers, biochemical markers (such as liver function, CRP, and electrolytes), and the TST-CT of patients infected with COVID-19 disease were assessed retrospectively (Tables 1–3). Two researchers assessed the collected data forms independently.

Statistical Analysis

The R 3.6.0 (www.r-project.com) was used to perform all of the statistical analyses. The Anderson–Darling test and Q-Q plots were used to check the normality of the variables. The homogeneity of the variances by group was examined using the Levene test. Data was

described as mean \pm standard deviation (range), median (interquartile range), and numbers (%) for the general characteristics of the patients with COVID-19. The Welch's t test, the Mann–Whitney U test, the χ^2 test with the Yates continuity correction, and the Fisher's exact test were used, as appropriate, to compare the general characteristics between the patient groups. Considering the possible extreme outliers under the pandemic conditions, the values for the laboratory findings were presented as trimmed mean (\pm SEM: standard error of mean), which was calculated with a 10% trim proportion and a robust estimator against the outliers. The Yuen's test (robust independent-samples t test) and the robust paired-samples t test were used to compare these findings. The comparisons were applied considering four situations: the LMWH and the control groups before treatment, the LMWH and the control groups on the seventh day of treatment, and the LMWH group before treatment and on the seventh day of the treatment (Figures 1 and 2). Finally, the calculated changes were compared by taking the difference of the seventh day and before the treatment in both groups; $p < 0.001$ was considered statistically significant. Univariate logistic regression analysis was utilized to view the risk factors. A multivariate logistic regression analysis was performed to evaluate the efficacy of the variant risk factors on the participant's discharge and scoring system; the odds ratio (OR) and 95% confidence interval (CI) were calculated (Table 4, 5). The forecasting value of the lymphocyte count, the D-dimer, and CRP, as assessed by the ROC curve and the cut-off value, which may predict the discharge, were identified afterward.

ROC Curve Analysis

The recovery performances of the laboratory parameters were evaluated by ROC analysis on the seventh day, as shown in Figure 3. The cut-off point for these parameters was determined according to the Youden index value. The risk factors included the lymphocyte counts, the level of CRP, and the D-dimer. The contributions of the risk factors were

determined based on the β value presented in Table 5. The area under the ROC curve for dividing the participants' LMWH, as compared to the control group, was applied for the threshold sensitivity, specificity, and accuracy. It was OR; 0.356 (standard error, 0.001; 95% CI, 0.089–0.674, β ; -1.032) for lymphocyte, OR; 0.974 (standard error, 0.001; 95% CI, 0.476–1.594, β ; -0.026) for D-dimer and OR; 0.628 (standard error, 0.001; 95% CI, 0.248–0.965, β ; -0.465) for CRP. The optimal threshold for identifying patients was 1.39, with 66.7% sensitivity and 64.6% specificity; 414, with 39.6% sensitivity and 85.4% specificity; and 14.3, with 58.3% sensitivity and 64.6% specificity, respectively.

Results

General Characteristics of the Patients with COVID-19

The LMWH group consisted of 15 males and 33 females aged between 40 and 68 years (average age 53.3 years), as shown in Table 1. The control group consisted of 18 males and 30 females aged between 44 and 66 years (average age 55.4 years). There was no substantial difference detected between the groups.

There were no significant differences between the groups in comorbidities such as diabetes mellitus, hypertension, coronary artery disease, gastrointestinal disease, cardiovascular disease, or other diseases. There were also no significant differences in COVID-19 pneumonia onset symptoms, with the inclusion of fever (temperature $\geq 37.4^\circ\text{C}$), dry cough, shortness of breath, sputum, myalgia, throat ache, nausea or vomiting, diarrhea, and headache. Similarly, there was no significant difference in the conventional therapy between the groups. These outcomes show that the general characteristics of the patient groups were both congruous and comparable.

Effect of LMWH on the Days to Conversion to Negative and the Length of the Hospital Stay of Patients with COVID-19

As shown in Table 2, the number of days to converting the virus to a negative outcome (time from the beginning of the stay in the hospital until the virus shedding) was 5.2 days (IQR 3.4 – 6.3) in the LMWH group and 7.6 days (IQR 6.5 – 9.7) in the control group ($p < 0.001$). A significant difference was detected between the groups. Also, the length of the stay in the hospital was 7.2 days (IQR 6.4 – 8.3) in the heparin group and 9.6 days (IQR 8.5 – 10.7) in the control group ($p < 0.001$). A significant difference was detected between the groups. **All of the participants demonstrated improvement after the treatment.**

Effect of LMWH on the Blood Routine in Patients with COVID-19

A significant difference was detected in the lymphocyte count between the groups before and after treatment, as shown in Table 2. **The after treatment lymphocyte count of the LMWH treatment group participants was elevated, and the detected difference was significant.** In addition, the changes in the lymphocyte counts in the LMWH group before and after therapy were significantly different according to the control group.

After treatment, the platelet results and levels were significantly elevated in the LMWH group, as compared to the control group. However, there was no significant difference detected between the groups in the monocyte, neutrophil percent, white blood cell (WBC), or hemoglobin levels.

Effect of LMWH on the Coagulation Function in Patients with COVID-19

The levels of the D-dimer and fibrin products prior to the application of LMWH in both groups were non-significant; however, these outcomes were significantly decreased after the LMWH treatment in the LMWH group, as compared to the control group, as shown in

Table 3. The patients' levels of D-dimer and FIB were significantly decreased in the LMWH group prior to and after treatment. The outcomes show that the application of LMWH improves the hypercoagulable state in COVID-19 patients. However, there was no significant difference detected in the PT and INR levels among the groups.

Effect of LMWH on the CRP in Patients with COVID-19

There were significant differences detected in the CRP levels between the groups both prior to and after the LMWH treatment, as shown in Table 3. Although the CRP levels were initially similar between the groups, these outcomes were significantly decreased in the LMWH group.

Effect of LMWH on the Cardiac Markers in Patients with COVID-19

As shown in Table 3, there were no significant differences detected in the creatine kinase isoenzyme B (CK-MB) levels between the groups prior to and after the LMWH treatment, and the results of both of the groups were decreased, as compared to prior to treatment. Similarly, there were no significant differences detected in the troponin-I levels between the two groups.

Effect of LMWH on the CT-SS in Patients with COVID-19

As shown in Table 3, there were significant differences detected in the CT-SS between the groups prior to and after the LMWH treatment. Although the CT-SS were initially similar between the groups, these outcomes decreased significantly after the LMWH treatment in the LMWH group, as compared to the control group.

Discussion

Cytokine storms are related to corruption in infectious illnesses, such as SARS and Middle East respiratory syndrome coronavirus (MERS); they are also a substantial cause of exacerbation in patients. Studies have revealed that heparin has some specifications other than anticoagulant properties. It performs anti-inflammatory effects by decreasing the extrication and biological efficacy of IL-6. However, the anti-inflammatory effects of heparin in the COVID-19 disease are not fully known. The main finding of this study is to evaluate the impact of the LMWH administration on the clinical course of the COVID-19 disease [12].

Although some studies have investigated the non-anticoagulant efficacy of LMWH, the real impact mechanism of LMWH remains unknown. This research investigated the anti-inflammatory effect of the LMWH molecule and its contribution to improvement in the COVID-19 disease. The LMWH is a heterogeneous molecule, and it has some features other than its anticoagulant effects. One of the effects of LMWH application is that it leads to a decline in inflammation. LMWH efficacy starts with inhibition of leukocyte transmigration stages into tissue. Heparin inhibits inflammation by its anticoagulant efficacy; these properties are closely intertwined. Some studies have investigated its anti-inflammatory effect. In an experimental model, Downing et al. found that LMWH reduced the inflammation and performed this efficacy without any anticoagulant effects. Ahmed et al. also showed that inhaled heparin has led to improvement in pulmonary functions and airway hypersensitivity in asthmatic patients [13,14].

Although the initial results were similar in the assay of distinctions of the lymphocyte counts, after the therapy period, the lymphocyte counts were more elevated in the LMWH group than the control group, which is consistent with Huang et al. This shows that LMWH

can elevate the lymphocyte counts and contribute to improvement of the disease. There are some causes for this. First, LMWH consists mainly of oligosaccharides, and it can be explained that short oligosaccharide chains, which are common in LMWH, may lead to the compression of cytokine storms; in addition, oligosaccharide chains, which are reproduced from nitrous acid depolymerization of LMWH, were able to bind to antithrombin-III (AT-III), and this indicates that the therapeutic effect for the hypersensitivity of LMWH is independent of the anti-coagulant impact [15–21].

One large trial about sepsis demonstrated that LMWH decreased the mortality rates. This data suggests that LMWH regressed the inflammation by its non-anticoagulant effects. Camprubi-Rimblas et al. revealed that LMWH may treat acute respiratory distress syndrome (ARDS) by improving lung inflammation. In addition, Paulsson et al. showed that heparin might heal a lung infection by struggling with heparan sulphate, which can cause a cytokine storm in COVID-19, by preventing the pathogens that bind host cells. However, the RASTEN study revealed that there was no significance detected in the survival time in lung cancer patients; although this was a disappointing outcome for LMWH, the dose of molecule in this study was prophylactic and the stage of the participants was higher; therefore these results are incongruous [22–24].

Societies have detected that age and comorbidity are reported as possible risk factors for patients who are infected with COVID-19. Moreover, some studies have demonstrated that thorax CT scans and laboratory markers, such as complete blood count, liver enzyme markers, coagulation parameters, inflammatory markers, and the quantity of immune cells of COVID-19 patients were connected with the severity of the illness [25–27].

In some studies on the COVID-19 disease, the D-dimer levels were increased substantially according to the disease severity. Tang et al. found that elevation of the D-dimer outcomes was correlated with mortality. Furthermore, Zhang et al. detected that elevated

results of D-dimer effects the hospital mortality, so it is shown as a helpful parameter for following up the improvement of COVID-19 patients. Therefore, D-dimer was used for an evaluation index marker for the progression of the illness in this research. The mean outcomes of D-dimer were higher in the LMWH group than the control group. This outcome is in line with the achievement of the LMWH performed group. The determination of the differences demonstrated that LMWH had better efficacy for lowering the D-dimer levels [11,28].

Novel studies have suggested that analyzing the CRP and lymphocyte count revealed the efficacy of treatment of COVID-19. When we touched on the differences between the groups, significant differences were detected in the decreasing of the CRP results and the elevation in the lymphocyte counts of the LMWH group, as compared to the control group [29,30].

In the study analysis of CRP outcomes, Walters et al. indicated that LMWH reduces CRP outcomes, indicating its anti-inflammatory effect. Furthermore, the ARMADA study revealed that inflammatory markers such as CRP were decreased in the heparin group. According to the control group, the CRP outcomes in the present study were lower in the LMWH group after the therapy period, which is in line with Walters et al. and the ARMADA study. This showed that LMWH can improve the CRP levels and contribute to improvement of the disease [31,32].

Novel studies also investigated some pathobiological perspectives, such as acquired thrombophilia in COVID-19, which were not evaluated previously and may enlighten future research. Patients with COVID-19 frequently have a higher risk for thrombotic situations. Therewithal, fibrin-based occlusions in microvessels have been found in the lung histopathology specimens examined in deceased COVID-19 patients [33–35].

Some synergistic mechanisms such as hypoxic vaso-occlusion, activation of cells by viral transduction, and cytokine storms, which have been detected in COVID-19 disease, may

lead to micro and/or macro thrombosis [36,37]. Furthermore, numerous studies have demonstrated that activated neutrophils conduce the spreading of thrombus, which affects vascular beds [38,39].

In accordance with these pathophysiological outcomes, some conventional gargling drugs suggested in the initial treatment of COVID-19 for leading the oropharyngeal viral shedding, such as Ankaferd hemostat, contain *Glycyrrhiza glabra* (which contributes to anti-inflammatory efficacy by way of a decline in the high mobility group box 1 protein [HMGB1] secretion) [40].

Through the logistic univariate regression model, this study detected that WBC outcomes, count of lymphocytes, neutrophils, D-dimer, FIB, CRP, and PLT were independent risk factors for participants. Moreover, some studies have shown that most of the patients infected with COVID-19 displayed with lymphocytopenia, elevated D-dimer levels, CRP, and in some cases elevated liver enzymes such as AST and ALT. Hematologic and biochemical outcomes were seriously elevated in patients who were followed up at ICU, recommending that the severity of the disease may have a relationship with cytokine storms and hyper inflammation. In addition, the logistic multivariate analysis model demonstrated that D-dimer, CRP levels, and lymphocyte counts were the main risk factors for the disease seriousness, which has a relationship with inflammation and hypercoagulation [41,42].

Although some scoring modules have more variables correlated with the disease prognosis, no scoring system has been accepted as a rule. Gong et al. formulated a seven-parameter system that included complete blood count and biochemical markers for identification of severity[43]. Chen et al. also performed a system for predicting the prognosis of the disease, including comorbidities, anti-inflammatory markers, and demographics [44]. The present study utilized the scoring system designed by Dong et al. for evaluating disease severity and assessing the treatment time by a simple formula, in comparison to other systems

that are confusing and may lead to misunderstandings [45]. Although the present system only contains three variables, it has better efficacy for distinguishing participants whose progress may turn to severity and respond to treatment conveniently. CRP was used as a scoring parameter, instead of the erythrocyte sedimentation rate (ESR), for evaluating anti-inflammatory efficacy more clearly [46]. In comparison to these studies, the present scoring system is a simple and rapidly detecting module for patients whose prognosis may become worse.

In addition to the scoring system, this research assessed the TST-CT outcomes for evaluating the improvement of patients and used it to assist in observing the improvement in the lungs. TST-CT is a sensitive apparatus and is better than chest x-rays for investigating the pulmonary parenchyma. Therefore, this method has become a pioneer diagnostic, and it is a helpful method for early detection of COVID-19 disease. The characteristic radiological outcome of COVID-19 is ground-glass opacities and/or consolidation asymmetrically located at peripheral lodges. The radiological manifestations are similar to those of SARS and MERS disease. The SARS and MERS disease both frequently show single lesions unilaterally and asymmetrically in the lungs. The TST-CT scores and the screened lesions were improved at the end of the follow-up period, according to the initial time outcomes [47,48].

This study had some limitations. First, it was a retrospective study, and the patient population was small; however, the sample size was considered adequate to draw relevant conclusions. The present study showed that research groups have an inclusive population treated in our center. Second, none of the patients who were treated in the ICU joined the study; all of the participants were discharged, and had no complications or mortality detected. Finally, the influence of other novel therapies on these patients was not evaluated. Because the research period was only over one week, it is possible that some non-pharmacological

changes were introduced in the management of patients as the medical societies learned more about this disease over time.

Conclusion

In conclusion, this research suggest that LMWH therapy added to conventional treatment has been shown to contribute to clinical and laboratory improvement in COVID-19. Those improvements might be the result of the anti-inflammatory effects of LMWH.

Disclaimers/Conflicts of Interest

The authors have no conflict of interest. They also declare that there was no funding source for conducting this study. They did not receive any funding regarding the current study.

Ethical Approval

The University Hospital of Medicine faculty approved the study (COVID-19–2020, No. 2020/037). This study was performed in accordance with the Helsinki Declaration of 1964 and its later amendments.

REFERENCES

1. Perlman S. Another Decade, Another Coronavirus. *The New England Journal Of Medicine* 2020;382: 760–762. doi: 10.1056/NEJMe2001126
2. World Health Organization (WHO), Clinical management of severe acute respiratory infection when novel coronavirus (2019-nCoV) infection is suspected: interim guidance, 28 January 2020
3. Xu XW, Wu XX, Jiang XG, Xu KJ, Ying LJ et al. Clinical findings in a group of patients infected with the 2019 novel coronavirus (SARS-Cov-2) outside of Wuhan, China: retrospective case series. *British Medical Journal* Version 2. *BMJ*. 2020 Feb 19; 368: 606-613. doi: 10.1136/bmj.m606
4. Liu Y, Yang Y, Zhang C, Huang F, Wang F et al. Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. *Science China Life* 2020 Mar;63(3):364-374. doi: 10.1007/s11427-020-1643-8
5. Chan MH, Wong VW, Wong CK, Chan PKS, Chu CM et al. Serum LD1 isoenzyme and blood lymphocyte subsets as prognostic indicators for severe acute respiratory syndrome. *Journal of Internal Medicine* 2004 Apr; 255(4):512-518. doi: 10.1111/j.1365-2796.2004.01323.x
6. Hou H, Zhang B, Huang H, Luo Y, Wu S et al. Using IL-2R/lymphocytes for predicting the clinical progression of patients with COVID-19. *Clinical Experimental Immunology*. 2020 Jul;201(1):76-84. doi: 10.1111/cei.13450
7. Wang D, Hu B, Hu C, Zhu F, Li X et al. Clinical characteristics of 138 hospitalized patients with 2019 novel coronavirus infected pneumonia in Wuhan, China. *Journal of The American Medical Association* 2020 Feb 7;323(11):1061-1069. doi: 10.1001/jama.2020.1585

8. Terpos E, Ntanasis-Stathopoulos I, Elalamy I, Kastritis E, Sergentanis TN et al. Hematological findings and complications of COVID-19, *American Journal of Hematology*. 2020 Jul;95(7):834-847. doi: 10.1002/ajh.25829
9. Tang N, Bai H, Chen X, Gong J, Li D et al. Anticoagulant treatment is associated with decreased mortality in severe coronavirus disease 2019 patients with coagulopathy, *Journal of Thrombosis and Haemostasis*. 2020 May;18(5):1094-1099. doi: 10.1111/jth.14817
10. Tang N , Li D, Wang X , Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia, *Journal of Thrombosis and Haemostasis*.2020 Apr;18(4):844-847. doi: 10.1111/jth.14768
11. Chang YC, Yu CJ, Chang SC, Galvin JR, Liu HM et al. Pulmonary sequelae in convalescent patients after severe acute respiratory syndrome: evaluation with thin-section CT. *Radiology* 2005 Sep;236(3):1067-1075. doi: 10.1148/radiol.2363040958
12. Tichelaar YI, Kluin-Nelemans JCH, Meijeret K. Infections and inflammatory diseases as risk factors for venous thrombosis. A systematic review. *Thrombosis and Haemostasis*; 2012 May;107(5):827-837. doi: 10.1160/TH11-09-0611
13. Downing LJ, Strieter RM, Kadell AM, Wilke CA, Greenfield LJ et al. Low-dose low-molecular-weight heparin is anti-inflammatory during venous thrombosis. *Journal of Vascular Surgery* 1998 Nov;28(5):848-854. doi: 10.1016/s0741-5214(98)70060-6
14. Ahmed T, Garrigo J, Danta I. Preventing bronchoconstriction in exercise-induced asthma with inhaled heparin. *New England Journal of Medicine* 1993 Jul 8; 329 (2): 90-95. doi: 10.1056/NEJM199307083290204

15. Li X, Wang L, Yan. S, Yang F, Xiang L et al. Clinical characteristics of 25 death cases infected with COVID-19 pneumonia: a retrospective review of medical records in a single medical center, Wuhan, China. *International Journal of Infectious Disease*. 2020 May;94:128-132. doi: 10.1016/j.ijid.2020.03.053
16. Li L, Tian H, Yong W, Zheng W, Yuan L et al. COVID-19 patients' clinical characteristics, discharge rate, and fatality rate of meta-analysis, *Journal of Medical Virology*. 2020 Jun;92(6):577-583. doi: 10.1002/jmv.25757
17. Huang C, Wang Y, Li X, Ren L, Zhao J et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet*. 2020 Feb 15;395(10223):497-506. doi: 10.1016/S0140-6736(20)30183-5
18. Ahmed T, Smith G, Vlahov I, Abraham WM. Inhibition of allergic airway responses by heparin derived oligosaccharides: Identification of a tetrasaccharide sequence. *Respiratory Research*, 2012 Jan 23;13(1):6. doi: 10.1186/1465-9921-13-6
19. Shastri MD, Stewart N, Eapen M, Peterson GM, Zaidi ST et al. Opposing effects of low molecular weight heparins on the release of inflammatory cytokines from peripheral blood mononuclear cells of asthmatics. *PLoS One*. 2015 Mar 4;10(3):e0118798. doi: 10.1371/journal.pone.0118798
20. Shastri MD, Stewart N, Horne J, Peterson GM, Gueven N et al. (2015). In-vitro suppression of IL-6 and IL-8 release from human pulmonary epithelial cells by non-

anticoagulant fraction of enoxaparin. *PLoS One*. 2015 May 11;10(5):e0126763. doi: 10.1371/journal.pone.0126763. eCollection 2015

21. Shastri MD, Stewart N, Horne J, Zaidi ST, Sohal S et al. Non-anticoagulant fractions of enoxaparin suppress inflammatory cytokine release from peripheral blood mononuclear cells of allergic asthmatic individuals. *PLoS One* 2015 Jun 5;10(6):e0128803. doi: 10.1371/journal.pone.0128803

22. Camprubi-Rimblas M, Guillamat-Prats R, Lebouvier T, Bringue J, Chimenti L et al. Role of heparin in pulmonary cell populations in an in-vitro model of acute lung injury. *Respiratory Research* 2017 May 10;18(1): 89-95. doi: 10.1186/s12931-017-0572-3

23. Paulsson M, Riesbeck K. How bacteria hack the matrix and dodge the bullets of immunity. *European Respiratory Review* 2018 Jun 27;27(148):180018. doi: 10.1183/16000617.0018-2018

24. Ek L, Gezelius E, Bergman B, Bendahl PO, Anderson H et al. Swedish Lung Cancer Study Group (SLUSG): Randomized phase III trial of low-molecular-weight heparin enoxaparin in addition to standard treatment in small-cell lung cancer: the RASTEN trial. *Annals of Oncology*, 2018 Feb 1;29(2):398-404. doi: 10.1093/annonc/mdx716

25. He JL, Luo L, Luo ZD, Lyu JX, Ng MY et al. Diagnostic performance between CT and initial real-time RT-PCR for clinically suspected 2019 coronavirus disease (COVID-19) patients outside Wuhan, China. *Respiratory Medicine*. 2020 Jul;168:105980. doi: 10.1016/j.rmed.2020.105980

26. Liu P, Tan XZ. 2019 Novel Coronavirus (2019-nCoV) Pneumonia. *Radiology* 2020 Feb 04;200257. doi:10.1148/radiol.2020200257
27. Chung M, Bernheim A, Mei X, Zhang N, Huang M et al. CT Imaging Features of 2019 Novel Coronavirus (2019-nCoV). *Radiology* 2020 Feb 4: 200230. doi: 10.1148/radiol.2020200230
28. Zhang L, Yan Q, Fan H, Liu X, Liu Z et al. D-dimer levels on admission to predict in-hospital mortality in patients with Covid-19. *Journal Of Thrombosis And Hemostasis: JTH*. 2020 Jun;18(6):1324-1329. doi: 10.1111/jth.14859
29. Mousavi S, Moradi M, Khorshidahmad T, Motamedi. M. Anti-Inflammatory Effects of Heparin and Its Derivatives: A Systematic Review. *Advances in Pharmacological Sciences*. 2015;507151. doi: 10.1155/2015/507151
30. Oldgren J, Fellenius C, Boman K, Jansson JH, Nilsson TK et al. Influence of prolonged dalteparin treatment on coagulation, fibrinolysis and inflammation in unstable coronary artery disease. *Internal Medicine*. 2005 Nov; 258(5): 420-427. doi: 10.1111/j.1365-2796.2005.01562.x
31. Walters DL, Ray MJ, Wood P, Perrin EJ, Bett JHN et al. High-dose tirofiban with enoxaparin and inflammatory markers in high-risk percutaneous intervention, *European Journal of Clinical Investigation*, 2010 Feb; 40(2): 139-147. doi: 10.1111/j.1365-2362.2009.02237.x

32. Montalescot G, Bal-dit-Sollier C, Chibedi D, Collet JP, Soulat T et al. Comparison of effects on markers of blood cell activation of enoxaparin, dalteparin, and unfractionated heparin in patients with unstable angina pectoris or non-ST-segment elevation acute myocardial infarction (the ARMADA study), *The American Journal of Cardiology*, 2003 Apr 15;91(8):925-930. doi: 10.1016/s0002-9149(03)00105-x
33. Tian S, Weidong H, Li N, Huan L, Haibo X et al. Pulmonary Pathology of Early-Phase 2019 Novel Coronavirus (COVID-19) Pneumonia in Two Patients With Lung Cancer. *Journal of Thoracic Oncology* 2020 May;15(5):700-704. doi: 10.1016/j.jtho.2020.02.010
34. Zhe X, Lei S, Yijin W, Jiyuan Z, Lei H et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome. *Lancet Respiratory Medicine* 2020 Apr;8(4):420-422. doi: 10.1016/S2213-2600(20)30076-X
35. Fox SE, Aibek A, Jack LH, Guang L, Brown JQ et al. Pulmonary and cardiac pathology in African American patients with COVID-19: an autopsy series from New Orleans. *Lancet Respiratory Medicine* 2020 Jul;8(7):681-686. doi: 10.1016/S2213-2600(20)30243-5
36. Zuo Y, Srilakshmi Y, Hui S, Kelsey G, Melanie Z et al. Neutrophil extracellular traps in COVID-19. *JCI Insight* 2020 Jun 4;5(11):e138999. doi: 10.1172/jci.insight.138999
37. Barnes BJ, Jose MA, Baxter-Stoltzfus A, Borczuk A, Cools-Lartigue J et al. Targeting potential drivers of COVID-19: Neutrophil extracellular traps. *Journal of Experimental Medicine* 217, 2020 Jun 1;217(6):e20200652. doi: 10.1084/jem.20200652
38. Kapoor S, Opneja A, Nayak L. The role of neutrophils in thrombosis. *Thrombosis Research* 2018 Oct;170: 87-96. doi: 10.1016/j.thromres.2018.08.005
39. Thalín C, Hisada Y, Lundström S, Mackman N, Wallén H. Neutrophil Extracellular Traps: Villains and Targets in Arterial, Venous, and Cancer-Associated Thrombosis.

Arteriosclerosis Thrombosis and Vascular 2019 Sep;39(9):1724-1738. doi: 10.1161/ATVBAHA.119.312463

40. Haznedaroğlu IC, Çelebier M. Turk J Med Sci Anti-infective and wound-healing pleiotropic actions of Ankaferd hemostat. Turkish Journal of Medical Sciences. 2020 Apr 19. doi: 10.3906/sag-2004-94

41. Fan BE, Chong VCL, Stephrene Seok Wei C, Gek Hsiang L, Kian Guan Eric L et al. Hematologic parameters in patients with COVID-19 infection. American Journal Of Hematology. 2020 Jun;95(6):E131-E134. doi: 10.1002/ajh.25774

42. Guan WJ, Zheng-Yi Ni, Hu Y, Liang WH, Chun-Quan O et al. Clinical Characteristics of Coronavirus Disease 2019 in China. The New England Journal Of Medicine. 2020 Apr 30;382(18):1708-1720. doi: 10.1056/NEJMoa2002032

43. Gong J, Ou J, Qiu X, Jie Y, Yaqiong C et al. A Tool to Early Predict Severe Corona Virus Disease 2019 (COVID-19) : A Multicenter Study using the Risk Nomogram in Wuhan and Guangdong, China. Clinical Infectious Diseases, 2020 Apr 16;ciaa443. doi: 10.1093/cid/ciaa443

44. Chen R, Liang W, Jiang M, Guan W, Zhan C et al. Risk factors of fatal outcome in hospitalized subjects with coronavirus disease 2019 from a nationwide analysis in China. Chest. 2020 Jul;158(1):97-105. doi: 10.1016/j.chest.2020.04.010

45. Dong Y, Zhou H, Li M, Zhang Z, Guo W et al. A novel simple scoring model for predicting severity of patients with SARS-CoV-2 infection Transboundary and Emerging Diseases. 2020 May 29;10.1111/tbed.13651. doi: 10.1111/tbed.13651
46. Qu PF, Li R, Xu C, Chai W, Li H et al. A Clinical Pilot Study to Evaluate CD64 Expression on Blood Monocytes as an Indicator of Periprosthetic Joint Infection. Journal of Bone and Joint Surgery: American volume. 2020 Apr 30. doi: 10.2106/JBJS.20.00057
47. Wu J, Wu X, Zeng W, Guo D, Fang Z et al. Chest CT Findings in Patients with Corona Virus Disease 2019 and its Relationship with Clinical Features. Investigative Radiology 2020 May; 55(5):257-261. doi: 10.1097/RLI.0000000000000670
48. Ooi GC, Khong PL, Müller NL, Yiu WC, Zhou LJ et al. Severe acute respiratory syndrome: temporal lung changes at thin-section CT in 30 patients. Radiology. 2004;230:836–844, doi: 10.1148/radiol.2303030853

Table 1. General characteristics of the patients infected with COVID-19

	LMWH Group	Control Group	<i>p</i>-value

	(n=48)	(n=48)	
Characteristics			
Age (years)	53.3 ± 15.6	55.4 ± 11.6	0.469
Gender			0.667
Female	15 (31.3)	18 (37.5)	
Male	33 (68.8)	30 (62.5)	
Comorbidity	29 (60.4)	28 (58.3)	0.718
Hypertension	17 (35.4)	18 (37.5)	0.617
Diabetes Mellitus	14 (29.2)	10 (20.8)	0.123
Coronary Artery Disease	7 (12.2)	5 (8.7)	0.831
Gastrointestinal Disease	1 (2.1)	2 (4.2)	0.512
Other Disease	6 (12.5)	3 (6.3)	0.486

Values were presented as mean ± standard deviation (min – max), median (min – max) or numbers (n) and percentages (%)

p-values were calculated by Welch's t test, Mann Whitney U test, χ^2 test with Yates continuity correction or Fisher's Exact test, as appropriate

Table 2. Signs, symptoms and clinical outcomes

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	LMWH Group (n=48)	Control (n=48)	p-value
Signs			
Fever (temperature $\geq 37^{\circ}\text{C}$)	38 (79.1%)	36 (75.0%)	0.808
Dry Cough	33 (68.8%)	30 (62.5%)	0.667
Shortness of breath	24 (50.0%)	28 (58.3%)	0.413
Sputum	13 (27.1%)	14 (29.2%)	0.825
Myalgia	10 (20.8%)	16 (33.3%)	0.251
Throat ache	8 (16.7%)	6 (12.5%)	0.772
Nausea or vomiting	1 (2.1%)	6 (12.5%)	0.135
Diarrhea	3 (6.9%)	4 (8.3%)	0.683
Headache	4 (8.3%)	2 (4.2%)	0.277
Time from initial symptoms to admission to hospital (days)	2.2(1.1-3.4)	2.4(1.6-3.2)	0.816
Time from hospitalization to viral shedding of disease (days)	5.2 (3.4 – 6.3)	7.6 (6.5 –9.7)	<0.001
Length of stay in hospital	7.2 (6.4 – 8.3)	9.6 (8.5 –10.7)	<0.001
Antibiotic therapy	48	48	0.997
azitromycin	38(79.1%)	39(81.2%)	
moxifloxacin	42(87.5%)	41(85.4%)	
Antiviral therapy			0.994
favipiravir	32(66.6%)	33(68.7%)	
Oseltamivir	43(89.5%)	42(87.5%)	

Response to treatment			
improved	48(100%)	30(62.5%)	0.212
steady	0	18(37.5%)	
disruptive	0	0	
CT-SS			
Score of left lung	5.0(4.0-6.0)	6.0(5.0-8.0)	<0.001
Score of right lung	5.0(3.75-6.0)	7.5(6.0-95)	<0.001
Total score	11.0(7.0-12.5)	13.5(12.5-16.0)	<0.001
CT-SS: CT severity score			

Table 3. Findings on laboratory findings and scores in the treatment period

	Initial Values			7 th Day of the Treatment			
	LMWH (n=48)	Control (n=48)	<i>p</i> ^a	LMWH (n=48)	Control (n=48)	<i>p</i> ^b	<i>p</i> ^c
WBC (k/uL)	9.26 ± 0.76	9.51 ± 0.80	0.497	6.40 ± 0.41	6.91 ± 0.37	0.365	0.435
Neutrophil (k/uL)	5.43 ± 0.54	4.88 ± 0.43	0.402	4.95 ± 0.45	4.84 ± 0.32	0.828	0.408
Monocyte (k/uL)	0.55 ± 0.05	0.52 ± 0.04	0.609	0.58 ± 0.03	0.65 ± 0.05	0.260	0.523
Monocyte percent (%)	7.39 ± 0.49	7.93 ± 0.64	0.491	7.87 ± 0.53	8.99 ± 0.67	0.175	0.318
Lymphocyte (k/uL)^{1,39}	0.77 ± 0.03	0.82 ± 0.02	0.094	1.39 ± 0.40	1.02 ± 0.03	<0.001	<0.001
Lymphocytes percent (%)	12.41 ± 1.06	13.66 ± 0.84	0.205	22.18 ± 2.02	19.86 ± 1.46	0.337	<0.001
HB (g/dL)	12.80 ± 0.35	13.29 ± 0.27	0.242	12.31 ± 0.31	12.96 ± 0.25	0.089	0.001
PLT (k/uL)	181.72 ± 11.83	189.05 ± 9.58	0.618	220.60 ± 11.17	229.15 ± 13.28	0.610	<0.001
ALT (U/L)	44.12±5.26	43.63±6.23	0.879	35.16±3.64	36.17±4.28	0.826	0.853
AST(U/L)	42.27±3.45	43.72±4.48	0.911	37.83±4.95	35.82±3.59	0.868	0.889
TB (mmol/l)	1.04±0.13	1.06±0.24	0.752	0.98±0.26	1.00±0.14	0.842	0.892
Na	138.25±3.45	141.40±2.48	0.764	142.26±3.25	144.52±4.28	0.815	0.795
K	4.32±1.13	4.46±1.48	0.827	4.53±1.26	4.61±2.01	0.874	0.855
Cre	1.10±0.25	1.16±0.44	0.745	0.98±0.55	1.01±0.62	0.825	0.794

CK-MB (ng/mL)	1.22 ± 0.15	1.28 ± 0.11	0.738	1.25 ± 0.15	1.33 ± 0.12	0.663	0.827
Troponin-I (ng/L)	9.46 ± 2.56	5.71 ± 0.79	0.152	9.06 ± 2.33	4.84 ± 0.99	0.089	0.750
PT (sn)	11.16 ± 0.15	13.36 ± 0.31	0.256	15.05 ± 0.13	11.78 ± 0.38	<0.001	<0.001
INR (INR)	1.03 ± 0.02	1.23 ± 0.03	0.540	1.43 ± 0.02	1.11 ± 0.04	0.069	0.229
D-DIMER (ng/mL)<414	815.02 ± 112.24	650 ± 59.68	0.182	414.10 ± 45.73	635.63 ± 39.96	<0.001	<0.001
CRP (mg/L)	43.66 ± 9.93	41.76 ± 7.20	0.134	14.32 ± 4.46	36.62 ± 3.09	<0.001	<0.001
FIB	588.24±10.25	595.35±12.36	0.627	326.35±12.25	550.37±15.38	<0.001	<0.001
CT-SS	15.0(12.0- 16.0)	15.3(13.5- 16.5)	0.983	11.0(7.0- 12.5)	13.5(12.5- 16.0)	<0.001	<0.001

Value s were presented as trimmed mean ± SEM (trimmed mean was calculated with %10 trim proportion) (SEM: standard error of mean)

p-values were calculated by Yuen's test (Robust independent samples *t*-test) and Robust paired samples *t*-test

p^a shows the comparison of LMWH and control groups in before the treatment

p^b shows the comparison of LMWH and control groups in 7th day of the treatment

p^c shows the comparison of before and 7th day of treatment in the LMWH group

Abbreviations: WBC: White blood cell, HB: Hemoglobin, PLT: Platelet, CK-MB: Creatinine kinase – myocardial band isoenzyme, PT: Prothrombin time, INR: International

normalized ratio, CRP: C reactive protein, FIB:Fibrinogen,, ALT: Alanine aminotransferase, AST: Aspartate aminotransferase ,TB: Total bilirubine, K:Potassium, Na: Sodium, Cre:Creatinine,

Table 4: Univariate logistic regression analysis of risk factors with COVID-19 improvement		
	Univariable OR (95% CI)	p value
Age	1.041 (1.018-1.057)	0.315
Gender	0.732 (0.457-1.104)	0.217
BMI (kg/m²)	0.538 (0.319-0.936)	0.472
CT-SS	12.30 (10.50-14.25)	<0.001
Laboratory findings		
WBC (k/uL)	2.485 (1.821-3.178)	<0.001
Neutrophils (k/uL)	1.514 (1.137-1.749)	<0.001
Lymphocyte (k/uL)	0.203 (0.015-0.322)	<0.001
Anti-Inflammatory Markers		
CRP (mg/L)	0.362 (0.131-0.926)	<0.001
D-DIMER (ng/mL)	0.657 (0.463-1.223)	<0.001
Fibrinogen	1.738 (1.176-2.549)	<0.001
CT-SS: CT severity score, CRP: C reactive protein		

Table 5: Multivariate logistic regression analysis of risk factors with COVID-19 improvement			
	β	OR (95% CI)	P
Lymphocyte (k/uL)	-1.032	0.356 (0.089-0.674)	<0.001
CRP (mg/L)	-0.465	0.628 (0.248-0.965)	<0.001
D-DIMER (ng/mL)	-0.026	0.974 (0.476-1.594)	<0.001

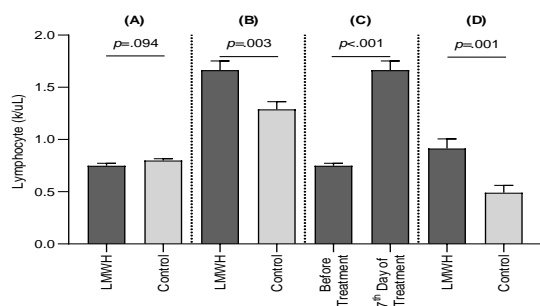


Figure 1. Effect of LMWH on Lymphocyte (k/uL) in the patients with COVID-19.

Data were expressed as mean \pm SEM. (A): Comparison of LMWH and control groups in before the treatment. (B): Comparison of LMWH and control groups in 7th day of the treatment. (C): Comparison of before and 7th day of the treatment in the LMWH group. (D): Comparison of the changes, which were calculated by taking difference 7th day and before the treatment in both groups.

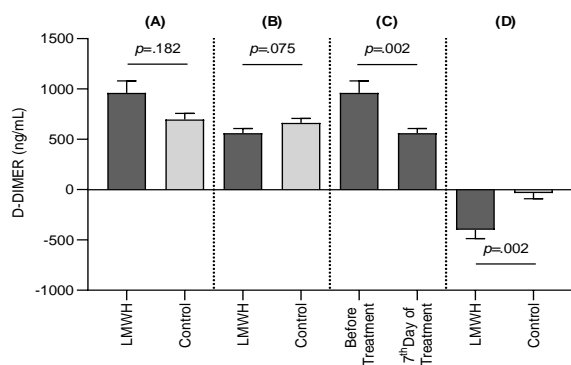


Figure 2. Effect of LMWH on D-DIMER (ng/mL) in the patients with COVID-19.

Data were expressed as mean \pm SEM. (A): Comparison of LMWH and control groups in before the treatment. (B): Comparison of LMWH and control groups in 7th day of the treatment. (C): Comparison of before and 7th day of the treatment in the LMWH group. (D): Comparison of the changes, which were calculated by taking difference 7th day and before the treatment in both groups.

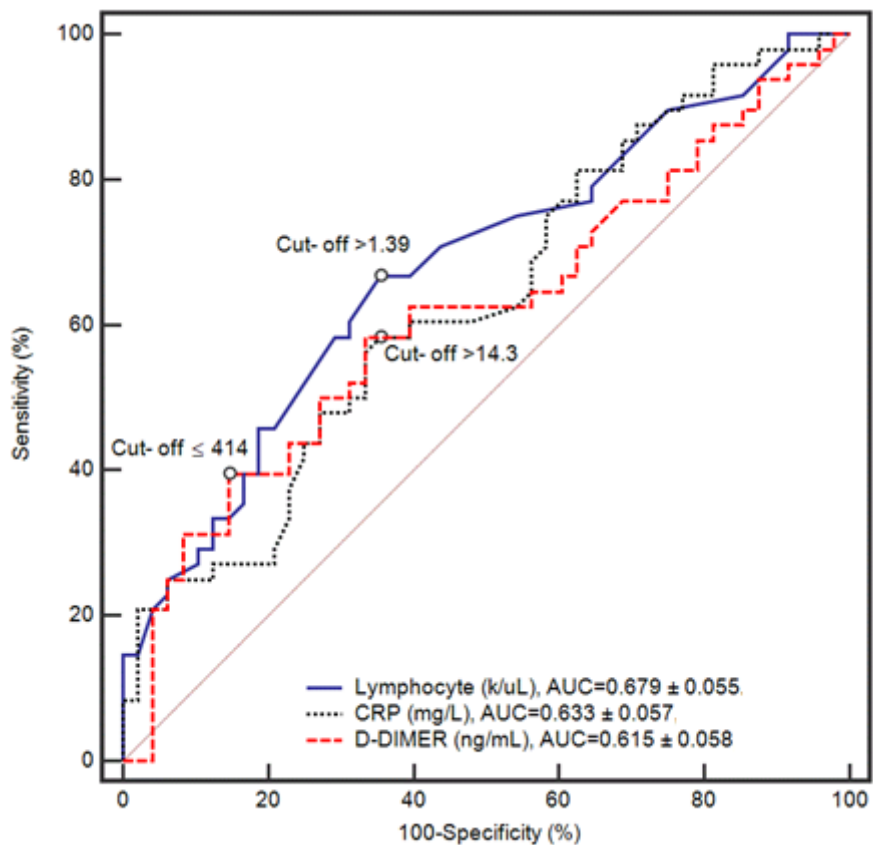


Figure 3: ROC curve for risk factors in patients infected with COVID-19. ROC = receiver operating characteristic.

