Understanding the immunological aspects of SARS-CoV-2 causing COVID-19 pandemic: A therapeutic approach

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pandemic: A therapeutic approach

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Abbreviations

ACE, angiotensin converting enzyme; Agn, angiotensin; AMs, alveolar macrophages, APCs,

antigen presenting cells; ARDS, acute respiratory distress syndrome; ASCs, antigen secreting

cells; AT2, alveolar type 2; BAL, bronchoalveolar lavage; CD, cluster of differentiation;

cDCs, conventional dendritic cells; CCL, C-C motif chemokine ligand; CDHR3, cadherin

related family member 3; CLpro, chymotrypsin-like protease; Covid-19, coronavirus disease,

2019; CTL, cytotoxic T lymphocyte; CTLA, cytotoxic T lymphocyte associated antigen;

CXCL, C-X-C motif chemokine ligand; CXCR, C-X-C motif chemokine receptor; DMVs,

dsRNA, double stranded RNA; E, envelope; FGF, fibroblast growth factor; G-CSF, granulocyte colony stimulating factor; GM-CSF, granulocyte-monocyte colony stimulating factor; GZMA, granzyme; HCoV, human coronavirus; H, hydrogen; HLA, human leucocyte antigen; ICIs, immune checkpoint inhibitors; IFN, interferon; Ig, immunoglobulin; IL, interleukin; IMs, inflammatory monocytes; IMMs, inflammatory monocyte-macrophages; IP-10, inducible protein 10; IRF, interferon regulatory factor; ISG, interferon stimulated gene; LAG3, lymphocyte activation gene 3; M, membrane; mABs, monoclonal antibodies; MBL, mannose binding lectin; MCP1, membrane cofactor protein 1; N. RS, middle east respiratory syndrome; MHC, major histocompatibility complex; MIP, macrophage inflammatory protein; moDCs, monocytes-derived dendritic cells; MSCs, nesenchymal stem cells; MYD88, myeloid differentiation primary response 88; N, nuclear psid; nABs, neutralizing antibodies; NGS, of next generation sequencing; NK cell, natural killer cells; NKG2A, NK group 2 member 2A; Nsps, non-structural proteins; ORTs, open reading frames; PAMPs, pathogen associated molecular patterns; PCR, solymerase chain reaction; PD1, programmed cell death protein 1; PD-L1, programmed cell deain protein ligand 1; PDGF, platelet derived growth factor; PLpro, papain like pro ea. ; Pp, polypeptides; PRRs, pattern recognition receptors; RA, rheumatoid arthritis; RBD, receptor binding domain; RBM, receptor binding motif; RLRs, RIG-I like recorders: RTC, replication transcription complex; S, spike; SARS-CoV-2, severe acute respiratory syndrome- coronavirus-2; scRNAseq, single cell RNA sequencing; SLE, systemic lupus erythematosus; ssRNA, single stranded RNA; STAT, signal transducer and activator of transcription; TIGIT, T cell immunoreceptor with Ig and ITIM domains; TIM 3, T cell immunoglobulin and mucin domain 3; TLRs, Toll like receptors; TMPRSS2, transmembrane protease serine 2; TNF, tumor necrosis factor; VEGF A, vascular endothelial growth factor A

Keywords: SARS-CoV-2; immune response; COVID-19 therapy.

2), a novel variant of coronavirus has recently emerged from Wuhan in China and has created havoc impulses across the world for a larger number of fatalities. At the same time studies are going on to discover vaccine against it or repurposing of approved drugs is widely adopted are under trial to eradicate the SARS-CoV-2 causing COVID-19. Reports have also shown that there are asymptomatic carriers of COVID-19 disease who can transmit the disease to others too. But the first line defense of the viral attack is body's strong and well-coordinated immune response producing excessive inflammatory innate reaction and impaired adaptive host immune defense may leading death upon the misfunctioning. Considerable works are going on to establish the relation between immune parameters and viral entry that might alter both the innate and adaptive immune system COVID patient by up riding a massive cytokines and chemokines secretion. This review mainly gives an account on how SARS-CoV-2 interact with our immune system and how doe: can immune system respond to it and along with that drugs which are being used can be used in fighting the disease and curative therapies as treatment for it has also been addressed.

1. Introduction

Evidences from the hartory focuses a spotlight on the incidences where coronavirus was found to the reason for the outbreak of disease and recently a new storm of coronavirus, named SARS-CoV-2 (Seter Acute Respiratory Syndrome-coronavirus-2), has been reported from Wuhan city, China. The disease is called Coronavirus disease, 2019 (COVID-19), named by WHO on Feb 11, 2020, that has first emerged in December 2019. As per the data, among the three outbreaks Middle East respiratory syndrome CoV (MERS-CoV) was the fatal most with a mortality rate of 34.77% while SARS-CoV stands out to be 10.87% of fatality and the SARS-CoV-2 has been reported to be 2.08% although is on a hike. Meanwhile the mortality rate in SARS-CoV-2 infection is lower than the previously reported two pandemics and its transmission rate is quite higher in comparison to the earlier ones [1]. As of September 4, 2020, there are 26,171,112 confirmed COVID-19 cases and death of

novel coronavirus known to infect humans. The severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is highly contagious [3]. The most prime transmission route of COVID-19 is droplets formation of aerosols including all other possible modes of direct contact. SARS-CoV-2 incubation period is approximately 5–14 days or 24 days in some cases as per the retrospective pandemic report identified [4]. Most of the patient suspected for COVID-19 positive require the supportive care, isolation to avoid the transmission chances and stronger immunity for recovery. Though the actual mechanism is still unclear, few anti-viral drugs such as remdesivir, lopinavir, ritonavir are used 1.5 treatment [5]. The SARS-CoV-2 infection in lung with adverse symptoms namely actual respiratory distress syndrome (ARDS) and which also the leads to severe lung injury mediated by immune system [6]. The SARS-CoV-2 positive lung biopsy report revealed that the content bilateral diffuse damage of alveoli, proliferation of fibroblast and activated circulated CD4+/CD8+ lymphocytes [7]. Due to the rapid global SARS-CoV-2 transmission, investigations are much needed for the development of effective immunotherapy. In view of this context, addressing of the immunological aspects of SARS-CoV-2 spreading COVID-19 become a major focus.

2. SARS-CoV background

According to WHO, SA SS-CoV-2 is from the beta lineage of the coronavirus family of group 2B with 70% or letic similarities with SARS-CoV [8]. There is four genera classification of family namely *Alphacoronavirus*, *Betacoronavirus*, *Deltacoronavirus* and *Gammacoronavirus* [9]. Cryo-electron tomography and cryo-electron microscopy gives an idea about the morphology of SARS-CoV. HCoV-229E and HCoV-OC43 are the two human coronavirus that are responsible for causing mild respiratory dysfunctions in humans before the rise of SARS-CoV infection in 2002, thereafter emergence of two new human coronavirus, HCoV-NL63 in 2004 and HCoV-HKU1 in 2005 has occurred, where HCoV-229E and HCoV-NL63 are found in bats [10]. Genome sequencing of SARS-CoV-2 revealed more or less 79% similarity with SARS-CoV and 50% similarity MERS-CoV according to genome sequencing that infected bat species underwent a series of genetic mutations and

the life risk situation that is prevailing all over the world because of SARS-CoV-2, raises a question forward regarding the origin of the virus. The most probable origin that has been brought to light is the zoonotic transfer of the virus from the illegally imported Malayan pangolins (Manis javanica) as genetic and evolutionary evidences suggest that the SARS-CoV harbored by these pangolins is 91.02% similar to the SARS-CoV-2 [12]. While angiotensin converting enzyme 2 (ACE2) host receptor sequence in bats (Rhinolophus sinicus), pangolins and human were taken under consideration, it revealed that the ACE2 sequence similarity between human and bats was 80.60% which is less similarity than between human and pangolins is 84.76%, indicatory that pangolins can be the original host or intermediate host of SARS-CoV-2 and therefore can promote transmission of the virus [13]. However, the genetic analysis of SARS-CoV-2 shows greater than 80% similarity compared with SARS-CoV and also more or e.s 50% similarity compared with MERS-CoV, both of them have a common origin to e. Lat [11]. According to phylogenetic analysis suggests that COVID-19, seventh member of the family of beta-coronavirus, is classified as a member of the ortho-coronavirina sunfamily and can be counted within the clade of the subgenus sarbecovirus [14]. R lacing to the previous epidemiological investigations we can figure out that the emergence of the new coronavirus is of zoonotic origin, keeping in mind the food habits of the Chares, people.

Being RNA virus, SARS-COV-2 has a high mutation rate that may involve in increasing virulence and pathogenecity of the infection in patients. Mutations in the surface proteins could change the tropism of the virus and increase its adaptability in new host with greater pathogenecity. Accumulation of mutations in SARA-CoV-2 may result in higher potency of pathogenecity. According to studies, high levels of mutations have been found in NSP and S proteins (Table 1). Current scenarios of COVID cases with 61.8M (million) cases and over 1.4M deaths globally, reported by WHO on 1st December 2020, shows a gradual spike in the COVID cases [15]. High level of mutations in S proteins may indicate a second wave of COVID19 with greater severity if essential steps are not taken.

The first five reported cases of COVID-19 in December, 2019 were hospitalized with ARDS and one deceased. Among all human associated CoVs, four patients were having mild respiratory symptoms, while two among them, with the infection of SARS-CoV and MERS-CoV were having severe respiratory diseases [24] which mainly had been transmitted from animals to humans via an intermediate mammalian host [25]. The results of next generation sequencing (NGS) or Real-time polymerase chain reaction (PCR) of patient's sputum targeted for the envelope gene of CoV confirmed the positive infection for COVID-19 [26] and SARS-CoV-2 shares almost 80% genome similarity with JARS-CoV [4]. Patients with positive infection of SARS-CoV-29, an enveloped single stranded RNA (ssRNA) virus with positive-sense RNA, show clinical manifestations [27]. In a nutshell, the pathogenesis of COVID-19 can be categorized as systemic disorders that include fever, dry cough, headache, fatigue, high sputum production, acute cardi c ii jury, dysponea, lymphopenia, cytokine storm and respiratory disorders that jucli de sore throat, sneezing, rhinorrhoea, severe pneumonia, ground-glass opacities, RNAaemia and ARDS. As per improvised current clinical symptoms, loss of smell and aste has become a new and potent symptom for COVID-19 along with other. A very recent study has shown that a higher expression of ACE2 and Type 2 transmembrane serine protease (TMPRSS2) on olfactory cells are highly affected by SARS-Co^V-2 resulting in the impairment of olfactory cells [28].

SARS-Cov-2, sourcing from symptomatic along with asymptomatic patients, after infecting a healthy person, has an incubation time of 4-14 days (average 3-7 days). Respiratory droplets from affected individual infect the healthy people to transmit the disease whereas it could also be transmitted through fecal-oral route because viral nucleic acid has been detected in the faeces and urine of COVID-19 patients. Along with the disease-causing comorbidities (cardiovascular, cerebrovascular, diabetes) and people of age more than 55 are showing more susceptibility to the COVID-19 infection but *Neeltje et al.* showed that cancer patients under chemotherapy and surgery treatment are more susceptible to SARS-CoV-2. On contrary, the patients who are receiving immunotherapy using immune checkpoint inhibitors

comparatively less prone to the COVID-19 disease [29].

4. Molecular mechanism of COVID-19 as pathogen

Based on the published literatures and the observations of the COVID-19 patients, the entry of the virus occurs via nasal and larynx mucosal membranes and reaches to the lungs via respiratory tract. S (spike) protein imparts virulence by binding to the host cell ACE2 receptor followed by their entry through clathrin-mediated endocytosis [30]. Different strains of Coronavirus can recognize different host cell receptors e.g. the receptor for SARS-CoV is ACE2 which affects the pneumatocytes (Type II) and cilia d bronchial epithelial cells [31,32], the receptor for HCoV-229E is aminopeptidase N or CD 3, the receptors for MERS-CoV is DPP4 (dipeptidyl peptidase4) or CD26 [32]. Ba ed on the genetic sequence analysis, difference lies between SARS-CoV-2 and SARS-CoV-1 and thus emerged as an absolute new betacorona virus of the novel corona virus of VID-19. Overall structural analysis of S protein between the two SARS-CoVs showed similarity of approximately 50 ~ 53% for the RBM (receptor binding motif), around 75% for the receptor binding domain (RBD) along with 76 to 78% whole protein [33]. As umption of using same receptors for binding comes from amino acid sequence analysis that revealed a high similarity in binding domain of ACE2 receptor in SARS-CoV [33,34]. In addition, ACE2 is an integral member of glycoprotein which is highly expressed in the lung, kidney, heart and epithelia of small intestine as well as endothelium [35]. The main function of ACE2 is the degradation of angiotensin (Ang)-II into Ang 1-7[35]. Pulmonary ACE2 maintains the balance between the circulating AngII/Ang1-7 levels. AngII, in response to hypoxia, induces pulmonary vasoconstriction and shunting in victims of lung injury and pneumonia is prevented [36].

In ARDS, ACE causes disease prognosis by increasing AngII level but ACE2 protects lung from failure by degrading AngII. Experimental evidences show that mice model where ACE2 is knockdown, drastic symptoms of ARDS is more prominent than wildtype while overexpression seems protective [37]. An increase of CD14⁺HLA⁻DR^{low} inflammatory monocytes (IMs) and Ficolin-1⁺ monocyte-derived macrophages has been detected by single

activation of interferon (IFN) signaling and monocytes recruitment decreases the alveolar potency and aids ARDS progression [38]. S protein on SARS-CoV-2 binds with greater affinity to host ACE2 receptors in comparison to SARS-CoV-1 [39]. Apart from ACE2⁺ cells, another study has focused on TMPRSS2, a cellular protease, which is required by the virus for entering into the cell as it helps the S protein on the virus surface to bind to the host ACE2 receptor, specially alveolar type-2 cells (AT2 cells) which express TMPRSS2 in large amount [40,41] whereas, the affinity towards the cadherin related family member 3 (CDHR3), a rhinovirus-C receptor at ciliated epithelial cells of the upper airway is still not cleared [42]. COVID-19 results into the inflammation in the king tissues due to less frequent exchange between oxygen and carbondioxide upon decrease in haemoglobin. This occurs due to the role of open reading frames (ORF1ab, ORF3a, ORF10), which attacks the 1-beta chain of haemoglobin into porphyrin ORF8 and sur are glycoproteins attaches. This mechanism can be treated with drugs like chloroqu'ne, favipiravir [43]. Transcriptomic study revealed that the genome of the virus is highly complicated and undergoes innumerable transcription events that in turn contributes to the production of unknown ORFs harboring mutations and undergoes recombination evert. Papid evolution of the virus, aids the virus to be drug resistant, frequently altered host specificity, thereby contributing to the virulence of the virus [44].

5. COVID-19 affecting immune system

5.1. Antigen presentation

Whenever a pathogen enters into our body, it is recognized as antigen and presented through antigen presenting cells (APCs) via major histocompatibility complex (MHC) molecules present on their surface. The exact mechanism of presentation of coronavirus is not fully known. According to the researches on SARS-CoV, MHC I molecule mediates the antigen presentation of the virus [45] and sometimes MHC II also participates in the process [46]. Previous data has shown a variety of polymorphisms such as human leukocyte antigen

infection susceptibility of SARs virus [46,47]. The protective alleles of HLAs mainly HLA-A*0201, HLA-DR0301 and HLA-Cw1502 [48]. Similarly, polymorphism like HLA-DRB1*11:01 and HLA-DQB1*02:0 in MHC II molecules elevates the risk for developing MERS-CoV infection. Moreover, mannose binding lectin (MBL) also present SARS-CoV to the immune cells [49]. The exact MHC type for the antigen presentation of COVID-19 disease is undeciphered but available data may shed some light in conducting further researches. The prospects that needed experimental focus is exact which type of MHC molecule are involved in the antigen presentation of SARS-CoV.

5.2. Immune evasion

In SARS-CoV-2, pattern recognition receptors (PKKs) activates the innate immune responses via extracellular and endosomal Toll- like receptors (TLRs) in concert with cytosolic RIG-I like receptors (RLRs) [50]. Fellowing the activation of PRRs, downstream signaling cascades stimulates the cytokin s production like Type I/III IFNs as defense against virus, tumor necrosis factor Opha (TNF-α), interleukins (IL-1,IL-6, IL-18) and other proinflammatory cytokines [51]. The complex signaling pathways involving myeloid differentiation primary response 88 (MYD88) produces type I IFNs and activate the transcription factor NF-B which induces the transcription and production of proinflammatory cytokines [5]. Type-I IFNs activate the downstream signal transducer and activator of transcription (STAT) proteins that catalyses generation of interferon stimulated genes (ISGs) coded antiviral proteins like IFN-induced protein with tetratricopeptide repeats-1. This phenomenon retards the replication of the virus in both neighboring and infected cells by activating an immune response against the virus. So, how COVID-19 can cause severe infection in patients? What are the immune escape strategies that are being adapted by the deadly virus? Evidences suggests that, not only SARS-CoV but MERS-CoV too produces double-membrane vesicles (DMVs) and avoid detection of their double stranded RNA (dsRNA) by host [53]. The nonspecific proteins 1 (Nsp1) of SARS-CoV repress the activation of IFN regulatory factor 3 (IRF3) and IRF7 and together with nsp3,

of IFN is blocked by accessory protein 4a of MERS-CoV upon interaction with double stranded DNA (dsDNA) directly [55]. Furthermore, studies have shown that in MERS-CoV infection ORF4a, ORF4b, ORF5 and membrane (M) proteins blocks transport of IRF3 into the nucleus and also inactivates IFNb promoter [56]. The gene expressions for antigen presentation are also downregulated after MERs-CoV infection. Thus, SARS-CoV and MERS-CoV has modified to escape from host immune surveillance. Efficient data is not available to support the theory that SARS-CoV2 also uses the same mechanism to avoid immune surveillance. Table 2 provides a comparative study or. 'he immunological functions played by structural proteins of the virus.

5.3. Innate immune system and SARS-CoV-2

First line of defense comes from the cells of innate immune system that include residential macrophages, conventional dendriti; ce'ls (cDCs), monocytes-derived dendritic cells (moDCs), granulocytes and natura' ki ler cells [74]. In any viral infection, the innate immune system relies on Type I in terferon (IFN) responses whose downstream cascade regulate the viral replication and in luce adaptive immune responses. However, nCOVID-19 may dampen the IFN Type-1 re-ponse to terminate the anti-viral response. According to studies, SARS-CoV directly aftest macrophages and T cells [75]. Recent research has shown that SARS-CoV-2 induces CD169⁺ tissue-resident macrophages to produce IL-6 which results into lymphocyte apoptosis via upregulation of Fas in human spleen and lymphnode [76]. According to the data of scRNAseq of COVID patients, there was expansion of CD14⁺IL-β⁺ monocytes [77] and IL-β associated inflammation in peripheral blood [78] and IMs, Ficolin-1⁺ monocyte-derived macrophages and tissue-resident reparative alveolar macrophages (AMs) in pulmonary tissues of severe condition [79]. In severe infection, lung macrophages express high IL-1B, IL6, TNF and chemokines like C-C motif chemokine ligand (CCL) 2, CCL3, CCL4 and CCL7, C-X-C motif chemokine ligand (CXCL) 9, CXCL10, CXCL11 but CXCL16, whose product binds C-X-C motif chemokine receptor (CXCR) 6 was more highly expressed in patients with moderate infection [79]. Moreover,

neutrophils though CCR1 and CXCR2 [79]. According to earlier data, SARS-CoV-1 infection resulted in an diverging phenotype of AM phenotype which limits the trafficking of DC and activation of T cell [80] and YM1⁺ FIZZ1⁺ alternatively activated macrophages increased hypersensitivity in airway, thus worsening the fibrosis by SARS-infection [81]. These mechanisms, in SARS-CoV-2, need more research focus. Recent research revealed that ACE2 and SARS-CoV-2 N protein is also present CD169⁺ macrophages of spleen and lymph node SARS-CoV-2 patients that are involved in production of IL-6 [82]. As mentioned earlier, SARS-CoV-2 undergoes the process of causing infection via ACE2 receptors but very low macrophage percentage in lungs express ACE2 receptors. So, the question arises that is there any other receptor present through which SAR'-Cov-2 is infecting immune cells? Evidences revealed reduced number of natural killer (NK) cells, in peripheral blood are positively correlated with COVID severity [83-8⁵]. In influenza infection, CXCR3 mediated NK cell infiltration [86]. In vitro study s'.ow; increased level of CXCR3 ligand (CXCL9-11) in SARS-CoV-2 infected tissue of human lung along with expanded monocyte level stimulated by CXCR3 ligand in S/R3-20V-2 infection [38]. These studies suggest that the CXCR3 pathway recruits NK (en in SARS-coV-2 infected patient the lungs from peripheral blood. Recent studies have shown that peripheral blood NK cells of SARS-coV-2 patients with deceased expression of enzymes such as granzyme B, granulysin and also reduced surface markers CD107a, Ksp37, and an impaired chemokine production of TNF-α and IFN-y that suggest an impaired cytotoxicity [84,87]. Moreover, SARS-CoV-2 infection has shown less number of CD16⁺KIR⁺ peripheral blood NK cells [88]. The expression of immune checkpoint NK group 2 member 2A (NKG2A) is increased with the upregulation of genes encoding inhibitory receptors lymphocyte activation gene 3 (LAG3) and T cell immunoglobulin and mucin domain 3 (TIM3) on NK cells of COVID patients [84,87]. Thus SARS-CoV-2 impairs the activity of NK cells. The impaired immune response stimulated by SARS-CoV-2 has been summarized in Fig 1.

Antigen presentation by APCs to other immune cells subsequently activates pathogen (virus) specific B cells and T cells. Similarly in SARS-CoV, viral infection a typical pattern of IgG and IgM has been observed where the IgM antibodies disappear in the 12th week but the IgGs which are viral S-specific and N-specific, last for a longer period [89]. The near-universal presence of IgGs, IgM, IgAs and neutralizing IgGs antibodies (nABs) in COVID patients indicates a humoral immune response mediated by increase in B cells. Covid-19 patients show higher levels of antigen secreting cells (ASCs) derived from precursor naïve B cells. These B cells are regarded as double negative2 (DN2) as 'bey lack naïve IgD, memory CD27 markers, CXCR5 and CD21 markers. The ASCs expressing levels of CD11c and T-bet molecular markers and respond to TLR7. Patients in early stages with high levels of ACEs provides a protective function against eradication of virus whereas in later stages high levels ACE show poor outcomes [90]. SARS-CoV-2 specific IgG of S protein was found in the serum of patient even after 60 days of s mprom onset, which decreased within 8 weeks of onset of post symptom [51]. Furtber studies are needed on the existence of viral specific IgG* memory cells in recovered CO VD patients.

Latest data has shown that in COVID-19 patients the peripheral count of CD4⁺ and CD8⁺ T cells have been greatly reduced but they were hyperactive in status. Additionally, a hike in highly proinfit metal ry CCR4⁺CCR6⁺ CD4⁺ T cells (Th17 cells) producing IL-17 and granulysin expressing Te cells were observed in patients with severe immune injury [91]. Moreover, the cytotoxic Tc cells (CD8⁺ T cells) were much higher in number e.g. 31.6% cells were perforin positive, 64.2% cells were granulysin positive and 30.5% cells were double positive for both perforin and granulysin [91]. These results implicated that the high number of Th17 and CD8⁺ T cells and their hyperactive function is responsible for severe immune inflammation in patients and produces low IFN-γ and TNF-α in CD4⁺ T cells and high granzyme B and perforin in CD8⁺ T cells in COVID-19 infected patients [92]. It has been reported that CD8⁺ T cells, developed during SARS-CoV infection, are specifically produced for the antigen S, M, E and N proteins. In SARS-CoV infection, CD8⁺ T cells have been

cells express CD45⁺CCR7⁺CD62L⁻ central memory T cells [93]. Th1 cells which was hyperactivated releases granulocyte-monocyte colony stimulating factors (GM-CSF) and IFNy. This recruits increased numbers of CD14⁺CD16⁺ monocytes that are inflammatory, stimulated by II-6 [94]. In moderately infected lung macrophages produced increased chemokines, that will attract T cells, via ethe engagement of CXCR3 and CXCR6 [38]. So, innate and adaptive immune cells interact with each other and they are involved in a positive loopback in expressing higher inflammation in COVID-19 infection. CD8⁺ T cells expressing high level cytotoxic genes such as granzyme K, A, B (GZMK, GZMA, GZMB) and XCL1 along with KLRC1 and were high in mild symptoms have been detected in bronchoalveolar lavage (BAL) of COVID patients [95]. Moreover, experimental analysis suggests that these memory cells lasts for 3-4 years after the infection has been cured and slowly diminishes in the absence of antigen after 4 years [93]. Moreover, all the subtypes of T cells found in SARS-CoV-2 infection, shows higher expression of negative immune checkpoint markers and exhaustion markers that is correlated with severe immune pathogenicity. The study of 10 patients group revealed increased levels of PD-1 in CD8⁺ T cells and Tim3 in CD4⁺ T cell were observed in three patients of both prodromal and symptomatic stages of SARS-CoV-2 infection [96]. Furthermore several other investigations reported increase in the expression of both co-stimulatory and inhibitory molecules such as OX-40 and CD137 [94], CTLA-4 and T cell immunoreceptor with Ig and ITIM domains (TIGIT) [92] and NKG2A [84], were found in T cells which suppressed cytotoxic activity. Till now there is no potent evidence of any memory cells developed in cured COVID-19 patients against SARS-CoV-2.

5.5. Cytokine storm and SARS-CoV-2

Till now according to the reports, the main cause of death due to COVID-19 is severe pneumonia and ARDS. The key cause behind the occurrence of ARDS is the severe "cytokine storm" in infected patients that resulted in pneumonia, respiratory failure and other organs failure. A high cytokine storm occurring in COVID-19 patients include IL1-β, IL1RA, IL7, IL8, IL9, IL10, basic-fibroblast growth factor 2 (FGF2), granulocyte colony stimulating

inflammatory protein 1α (MIP1α), MIP1-β, platelet derived growth factor B (PDGF-B), TNFα, and vascular endothelial growth factor A (VEGFA) [26,97]. The ICU patients show high levels of pro-inflammatory cytokines such as IL2, IL7, IL10, G-CSF, IP10, MCP1, MIP1α, and TNFα, which are positively correlated with disease severity [26]. In a report from Wuhan where 99 cases had been studied, an increase in total neutrophils, decrease in total lymphocytes and increased in serum IL-6 has been observed. A delayed IFN-I signaling was observed which accumulate inflammatory monocyte-macrophages (IMMs). This resulted in high levels of cytokine and chemokine in lung, vascular leakas, and impaired the response of viral-specific T cell [98]. In SARS infected patients, an elavated level of IL-6, IL-8 and inducible protein 10 (IP-10) has been found in lu. g ussue [99]. Increased levels of proinflammatory cytokines is mainly responsible for cavore lung injury, leading to demise of nCovid victims [91]. High levels of IP10 was related with immune mediated severe lung injury and apoptosis of lymphocytes ir S_F RS [99]. Together with the cytokines, certain chemokines such as CXCL10, IP1° CCL2, CCL3, CCL5, CXCL8, CXCL9 support the impaired systemic inflammatory response in SARS-CoV-2 [100]. In comparison with SARS-CoV, SARS-CoV-2 upregulate 1 nive chemokines namely CXCL1, CXCL5, CXCL10, CCL2 and IL-6 [101]. SARS-CoV 2 patients with more severe pneumonia and pulmonary syndrome showed correlated Light expression of GM-CSF⁺ and IL-6⁺CD4⁺ T cells, higher coexpression of IFN-y and GM-CSF in pathogenic Th1 cells, much higher expression of CD14⁺CD16⁺ monocyte [94]. In a nutshell, high infiltration of all type of immune cells such as T cells, monocytes, macrophages, NK cells, DCs, and secretion of their proinflammatory cytokines into lung cause severe ARDS leading to death in the patients.

6. Ongoing therapies

Currently there is no clinically approved antiviral vaccine for the treating SARS-CoV2. All patients are treated with supportive treatment strategies targeted to culminate the patients' symptoms (like pneumonia, fever, breathing problems) and often supported with combination of drugs. However, these strategies cannot be implemented for a long time.

other RNA viruses such as the Human Immunodeficiency Virus (HIV). Clinical trials are currently undergoing with combinational drugs mainly ritonavir and lopinavir. Several other drugs are under clinical trials such as Kevzara, a rheumatoid arthritis (RA) drug that decreases lung complications. Kevzara has been successfully tested in COVID-19 patients. As per data, there are 11 phase 4, 36 phase 2 and 4 phase 1 trails [102]. Table 3 encloses a list of commonly used combinational drugs for the treatment of COVID-19.

As it has been an absolute outbreak and pandemic disease declared by WHO, a specific cure has to be found out to cure the disease completely. According to the genomic and structural analysis of SARS-CoV-2, there are a number of therapeutic targets which are under clinical trials in different laboratories across the whole world.

6.1. Viral targets

The Washington Department of Hearth Administration has first introduced remdesivir which inhibit RNA dependent RNA polymerase activity intravenously and found that it has a potential to protect from SARS-CoV-2 infection. The combination of remdesivir and choloroquine has shown to prevent SARS-CoV-2 infection *invitro*. Therefore, other nucleotide analogue such as favipiravir, ribavirin can also be administered as potential inhibitors. There are certain proteases such as 3 chymotrypsin-like protease (CLpro) along with papain like protease (PLpro) that cleave viral polyproteins, can be the noble drug targets for the treatment. These also affects the replication of virus and antagonize IFN, IL-6. As SARS-CoV-2 binds with the ACE2 receptors of host cell, therefore targeting the S protein on the surface of the virus or the binding of the S protein and ACE2 can be a potential therapeutic target to combat COVID-19 infection. Fig 2 points out the role of various drugs in distinctive stages of SARS-CoV-2 replication process.

6.2. Antibody and plasma therapy

According to studies the development of recombinant monoclonal antibody (mAb)

human mAb, can bind with the RBD of SARS-CoV-2and can be used as candidate vaccine for SARS-CoV-2. Other mAbs, such as m396, CR3014, can be alternative against SARS-CoV-2. Recently, a recombinant mAb named tocilizumab has come into application that can bind to IL-6 receptor, thereby terminating its signal transduction but its efficiency is still under study [29]. In addition to it, virus neutralizing antibody isolated from convalescent serum of COVID-19 patients, who has recovered from the infection, is also administered in susceptible individuals as it proved to be promising treatment approach during the previous corona outbreaks. It can impose immediate immune response in the unaffected susceptible individuals [109]. The generation of antibodies against the proteins of the virus is being followed by Moderna Inc., MA, USA. There is also have for development of new mAbs, which may take less time to be available to the docume due to their speedy trials and their high specificity.

6.3. Development of Vaccine

In this pandemic situation, approved vaccines against SARS-CoV-2 are essentially required as soon as possible for decreasing disease severity together with reduced shedding and transmission of virus. Currently, no approved vaccine is available in the market to cure this disease. In the world full of darkness, a keen ray of light has been illuminated by the recent development of a vaccine mRNA1273 by The National Institute of Allergy and Infectious Diseases (NIAID), part of the National Institutes of Health (NIH). This mRNA1273 is now under phase one trial. The vaccine may be available in the near future, but the time require for this to reach the market depends on the efficacy and success in all three phases of clinical trials.

6.4. Epitope Mapping

Significant studies have been focused on identifying various target epitopes mapping on SARS-CoV-2 for development of targeted vaccines. Apart from antibodies

fragments (epitopes). This investigation has utilized the data on genetic differences and similarities among the three strains of coronavirus by utilizing bioinformatics analysis [110,111]. By implementing immune informatics scientists discovered five cytotoxic T lymphocytes (CTLs) epitopes and eight B-cell epitopes in the viral surface glycoproteins. Among the B cell epitopes three are sequential and rest five are discontinuous. Furthermore, CTL epitopes are activated as judged by molecular dynamicity that the interaction between the CTL epitopes and HLA chains of MHC-I complexes are mediated by hydrogen (H) bonds and salt bridges, indicating their efficacy to confer imm. The responses [111]. Another study has identified five linear and two conformational B 22.1 epitopes of SARS-CoV-2 surface proteins [112].

6.5. Stem cell therapy

This noble therapeutic approach and soft undertaken into study until two distinct studies conducted by China and detected that stem cell therapy can be a new aspect in the treatment of COVID-19. Intravenous a fusion of mesenchymal stem cells (MSCs) can play a vital role in curing the dysfunctions of the lung i.e. complications like pulmonary edema, dysfunction of air-exchange, AkDS, acute cardiac injury which are the results increased inflammation due to the account of the virally triggered cytokine storm caused damage to lung tissues [41,113]. These MSCs, depending on their properties of modulating the immune system and regeneration or differentiating capability can counteract the increased release of cytokines and their repairing capacity thus restore the damaged tissues, followed by curing the disease. Moreover, RNA-sequencing of transplanted MSCs has revealed the presence of undifferentiated transfused MSCs and remained to ACE2 negative thus there exist no chance for the virus to affect these cells [113].

7. Ongoing interventional studies

is undertaken across the world on observing the severity of COVID-19. From the increasing number of fatalities and affected individuals reported across the world, it is clear that COVID-19 is escaping the treatment strategies undertaken to eradicate it. Due to the immense capability of SARS-CoV-2 to mutate very rapidly, a challenge has been thrown to the scientists to develop a potent vaccine that can destroy it all. Keeping pace with the abovementioned treatment strategies, several drugs, which are used for treating other diseases, under clinical trial to find a solution to the menace of COVID-19. A list of interventional drugs and vaccines under trial has been illustrated in the Table 4.

Another concept of controlling the rapid spread on the virus is to develop herd immunity which is defined as decrease in population of susceptible individuals below the threshold value required for transmission. The contagions state of SARS-CoV-2 (R₀) varies between 2%-3%. So, for acquiring herd immunity the threshold value is 67% for this virus.

8. Conclusion

The current scenario of rapicly spreading and unpredicted infectious nature of SARS-CoV-2 demands an urgency to focus on basic science and clinical research. Though there are a few resemblances of immunovathogenesis of SARS-CoV-2 with SARS-CoV-1 and MERS but the differences are prominent enough to focus on new therapeutic targets for developing vaccines. Within short time there is significant knowledge about the immunology of SARS-CoV-2 infection which can aid in potent vaccine development. The emerging cases of asymptomatic situations is demanding a better and deep evaluation about the mechanisms of immune response following SARSCoV-2 infection to develop a promising therapeutic approach. Here, we reviewed some recent literatures that interrogate the viral entry, invasion, escape and immune mechanisms, the dysfunctions of various immune cells like T cells, NK cells, monocytes lineages with a brief view on the memory cells. We also addressed antibody and plasma therapy as well as vaccine development against SARS-CoV-2. Further studies are needed to explain the immune response varying in victims encompassing both affected and

accomplish the unmet needs.

Authors contributions

All authors have contributed equally in this work.

Conflict of interest

Authors do not have any conflict of interest.

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References

- [1] S.A. Meo, A.M. Alhowikan, T. Al-Khlaiwi, I.M. Meo, D.M. Halepoto, M. Iqbal, A.M. Usmani, W. Hajjar, N. Ahmed, Novel comparison 2019-nCoV: prevalence, biological and clinical characteristics comparison with SARS-CoV and MERS-CoV., Eur. Rev. Med. Pharmacol. Sci. 24 (2020) 2012–2019. https://doi.org/10.26355/eurrev_202002_20379.
- [2] WHO Coronavirus Disease (COVID-19) Dashboard | WHO Coronavirus Disease (COVID-19) Dashboard, (n.d.).
- [3] A.E. Gorbalenya, S.C. Baker, R.S. Baric, R.J. de Groot, C. Drosten, A.A. Gulyaeva, B.L. Haagmans, C. Lauber, A.M. Leontovich, B.W. Neuman et al. The species Severe acute respiratory syndromerelated coronavirus: classifying 2019-nCoV and naming it SARS-CoV-2, Nat. Microbiol. 5 (2020) 536–544. https://doi.org/10.1038/s41564-020-0695-z.

- characterisation and epidemiology of 2019 novel coronavirus: implications for virus origins and receptor binding, Lancet. 395 (2020) 565–574. https://doi.org/10.1016/S0140-6736(20)30251-8.
- [5] T. Pillaiyar, S. Meenakshisundaram, M. Manickam, Recent discovery and development of inhibitors targeting coronaviruses, Drug Discov. Today. 25 (2020) 668–688. https://doi.org/10.1016/j.drudis.2020.01.015.
- [6] M. Zhou, X. Zhang, J. Qu, Coronavirus disease 2019 (COVID-19): a clinical update, Front. Med. 14 (2020) 126–135. https://doi.org/10.1007/s11684-020-0767-8.
- [7] S. Tian, Y. Xiong, H. Liu, L. Niu, J. Guo, M. Liao, S.-Y. Xioo, Fathological study of the 2019 novel coronavirus disease (COVID-19) through postmortem care biopsies, Mod. Pathol. 33 (2020) 1007–1014. https://doi.org/10.1038/s41379-020-0536-x.
- [8] D.S. Hui, E. I Azhar, T.A. Madani, F. Ntourni, R. Kock, O. Dar, G. Ippolito, T.D. Mchugh, Z.A. Memish, C. Drosten, *et al.* The continuity 2019-nCoV epidemic threat of novel coronaviruses to global health The latest 2019 novel coronavirus outbreak in Wuhan, China, Int. J. Infect. Dis. 91 (2020) 264–266. https://doi.org/10.1016/j.ijid.2020.01.009.
- [9] S. Perlman, J. Netland, Coronaviruses post-SARS: Update on replication and pathogenesis, Nat. Rev. Microbiol. 7 (2009) 439–450. https://doi.org/10.1038/nrmicro2147.
- [10] T.S. Fung, D.X. Liu, Human Coronavirus: Host-Pathogen Interaction, Annu. Rev. Microbiol. 73 (2019) 529–557. https://doi.org/10.1146/annurev-micro-020518-115759.
- [11] J. Cui, F. Li, Z.L. Shi, Origin and evolution of pathogenic coronaviruses, Nat. Rev. Microbiol. 17 (2019) 181–192. https://doi.org/10.1038/s41579-018-0118-9.
- [12] T. Zhang, Q. Wu, Z. Zhang, Probable pangolin origin of SARS-CoV-2 associated with the COVID-19 outbreak, Curr. Biol. 30 (2020) 1346-1351.e2. https://doi.org/10.1016/j.cub.2020.03.022.
- [13] P.H. Guzzi, D. Mercatelli, C. Ceraolo, F.M. Giorgi, Master Regulator Analysis of the SARS-CoV-2/Human Interactome, J. Clin. Med. 9 (2020) 982. https://doi.org/10.3390/jcm9040982.

- novel coronavirus from patients with pneumonia in China, 2019, N. Engl. J. Med. 382 (2020) 727–733. https://doi.org/10.1056/NEJMoa2001017.
- [15] World Health Organisaton, COVID-19 Weekly Epidemiological Update Global summary, (2020) 1–22.
- [16] J.A. Plante, Y. Liu, J. Liu, H. Xia, B.A. Johnson, K.G. Lokugamage, X. Zhang, A.E. Muruato, J. Zou, C.R. Fontes-Garfias, et al. Spike mutation D614G alters SARS-CoV-2 fitness, Nature. (2020). https://doi.org/10.1038/s41586-020-2895-3.
- [17] S. Isabel, L. Graña-Miraglia, J.M. Gutierrez, C. Bundalovic Terma, H.E. Groves, M.R. Isabel, A.R. Eshaghi, S.N. Patel, J.B. Gubbay, T. Poutanen, *et al.* Evolutionary and structural analyses of SARS-CoV-2 D614G spike protein mutation now documented worldwide, Sci. Rep. 10 (2020) 1–9. https://doi.org/10.1038/s41598-020-70827-z.
- [18] S. Angeletti, D. Benvenuto, M. Bianchi, V. Giovanetti, S. Pascarella, M. Ciccozzi, COVID-2019: The role of the nsp2 and nsp3 in its pathogenesis, J. Med. Virol. 92 (2020) 584–588. https://doi.org/10.1002/jmv.25719.
- [19] T.A. Brief, Rapid increase of a SARS-CoV-2 variant with multiple spike protein mutations observed in the United Kingdom, (2020).
- [20] M. Pachetti, B. Marini, F. Benedetti, F. Giudici, E. Mauro, P. Storici, C. Masciovecchio, S. Angeletti, M. Ciccozzi, R.C. Gallo, *et al.*, Emerging SARS-CoV-2 mutation hot spots include a novel RNA-dependent-RNA polymerase variant, J. Transl. Med. 18 (2020) 1–9. https://doi.org/10.1186/s12967-020-02344-6.
- [21] R.A. Khailany, M. Safdar, M. Ozaslan, Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19.

 The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information, Gene Reports. 19 (2020) 1–6.

- B. Lee, Y.S. Goh, *et al.* Effects of a major deletion in the SARS-CoV-2 genome on the severity of infection and the inflammatory response: an observational cohort study, Lancet. 396 (2020) 603–611. https://doi.org/10.1016/S0140-6736(20)31757-8.
- [23] L.A. Holland, E.A. Kaelin, R. Maqsood, B. Estifanos, L.I. Wu, A. Varsani, R.U. Halden, B.G. Hogue, M. Scotch, E.S. Lim, An 81-Nucleotide Deletion in SARS-CoV-2 ORF7a Identified from Sentinel Surveillance in Arizona (January to March 2020), J. Virol. 94 (2020). https://doi.org/10.1128/jvi.00711-20.
- [24] V.M. Corman, J. Lienau, M. Witzenrath, Coronaviren als Ur ache respiratorischer Infektionen, Internist (Berl). 60 (2019) 1136–1145. https://doi.org/10.1637/c00108-019-00671-5.
- T.P. Sheahan, A.C. Sims, R.L. Graham, V.D. Menacı, Y. L.E. Gralinski, J.B. Case, S.R. Leist, K. Pyrc, J.Y. Feng, I. Trantcheva, *et al.* Broad-spe Y. un antiviral GS-5734 inhibits both epidemic and zoonotic coronaviruses, Sci. Transl. Med. 9 (2017) eaal3653. https://doi.org/10.1126/scitranslmed.aa/3653.
- [26] C. Huang, Y. Wang, X. Li, L. Ren, J. Zhao, Y. Hu, L. Zhang, G. Fan, J. Xu, X. Gu, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China, Lancet. 395 (2020) 497–506. https://doi.org/10.1015/S0140-6736(20)30183-5.
- [27] H.A. Rothan, S.N. Byrare Jdy, The epidemiology and pathogenesis of coronavirus disease (COVID-19) outbreak, J. Autoimmun. (2020) 102433. https://doi.org/10.1016/j.jaut.2020.102433.
- [28] K. Bilinska, P. Jakubowska, C.S. Von Bartheld, R. Butowt, Expression of the SARS-CoV-2 Entry Proteins, ACE2 and TMPRSS2, in Cells of the Olfactory Epithelium: Identification of Cell Types and Trends with Age, ACS Chem. Neurosci. 11 (2020) 1555–1562. https://doi.org/10.1021/acschemneuro.0c00210.
- [29] M. Bersanelli, Controversies about COVID-19 and anticancer treatment with immune checkpoint inhibitors, Immunotherapy. 12 (2020) 269–273. https://doi.org/10.2217/imt-2020-0067.

- cells through a novel clathrin- and caveolae-independent endocytic pathway, Cell Res. 18 (2008) 290–301. https://doi.org/10.1038/cr.2008.15.
- [31] W. Li, M.J. Moore, N. Vasilieva, J. Sui, Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus, Nature. 426 (2003) 450–4. https://doi.org/https://doi.org/10.1038/nature02145.
- [32] G. Lu, Y. Hu, Q. Wang, J. Qi, F. Gao, Y. Li, Y. Zhang, W. Zhang, Y. Yuan, J. Bao, *et al.* Molecular basis of binding between novel human coronavirus MERS-CoV and its receptor CD26, Nature. 500 (2013) 227–31. https://doi.org/10.1038/nature12328.
- [33] Y. Wan, J. Shang, R. Graham, R.S. Baric, F. Li, Receptor Recognition by the Novel Coronavirus from Wuhan: an Analysis Based on Decade-Long Structural Studies of SARS Coronavirus, J. Virol. 94 (2020) e00127-20. https://doi.org/10.1128/jvi.00127-20.
- [34] P. Zhou, X. Lou Yang, X.G. Wang, B. Hu, L. Zhang, W. Zhang, H.R. Si, Y. Zhu, B. Li, C.L. Huang, et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin, Nature. 579 (2020) 270–273. https://doi.org/10.1038/s^1586-020-2012-7.
- [35] C. Tikellis, M.C. Thomas, Angic ensin-Converting Enzyme 2 (ACE2) Is a Key Modulator of the Renin Angiotensin System in Health and Disease, Int. J. Pept. 2012 (2012) 256294. https://doi.org/10.1155/2012/256294.
- [36] D.G. Kiely, R.I. Cargill, N.M. Wheeldon, W.J. Coutie, B.J. Lipworth, Haemodynamic and endocrine effects of type 1 angiotensin II receptor blockade in patients with hypoxaemic cor pulmonale, Cardiovasc. Res. 33 (1997) 201–208. https://doi.org/10.1016/s0008-6363(96)00180-0.
- [37] Y. Imai, K. Kuba, S. Rao, Y. Huan, F. Guo, B. Guan, P. Yang, R. Sarao, T. Wada, H. Leong-poi, *et al.* Angiotensin-converting enzyme 2 protects from severe acute lung failure, Nature. 436 (2005) 112–6. https://doi.org/10.1038/nature03712.
- [38] M. Liao, Y. Liu, J. Yuan, Y. Wen, G. Xu, J. Zhao, L. Cheng, J. Li, X. Wang, F. Wang, et al. Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19, Nat. Med. 26 (2020)

- [39] K. Kuba, Y. Imai, S. Rao, H. Gao, F. Guo, B. Guan, Y. Huan, P. Yang, Y. Zhang, W. Deng, *et al.* A crucial role of angiotensin converting enzyme 2 (ACE2) in SARS coronavirus induced lung injury, Nat. Med. 11 (2005) 875–879. https://doi.org/10.1038/nm1267.
- [40] L. Mousavizadeh, S. Ghasemi, Genotype and phenotype of COVID-19: Their roles in pathogenesis, J. Microbiol. Immunol. Infect. (2020). https://doi.org/10.1016/j.jmii.2020.03.022.
- [41] A.A. Ali Golchin, Ehsan Seyedjafari, Ali Golchin 1, Ehsan Seyedjafari 2, Mesenchymal Stem Cell Therapy for COVID-19: Present or Future, Stem Cell Ke^{-y}. Reports. 16 (2020) 427–433. https://doi.org/10.1007/s12015-020-09973-w.
- [42] T.F. Griggs, Y.A. Bochkov, S. Basnet, T.R. Pasic, R. A. Brockman-Schneider, A.C. Palmenberg, J.E. Gern, Rhinovirus C targets ciliated airway extherial cells, Respir. Res. 18 (2017) 84. https://doi.org/10.1186/s12931-017-0567-0.
- [43] W. Liu, H. Li, COVID-19: Attacks the 1-Beta Chain of Hemoglobin and Captures the Porphyrin to Inhibit Human Heme Metabolism, ChemRxiv. Preprint (2020). https://doi.org/10.26434/chemrxiv. 11938173.v5.
- [44] D. Kim, J. Lee, J. Yang, J.W. Kim, V.N. Kim, H. Chang, The architecture of SARS-CoV-2 transcriptome, (2020) 1-25.
- [45] J. Liu, P. Wu, F. Gao, J. Qi, A. Kawana-tachikawa, J. Xie, C.J. Vavricka, A. Iwamoto, T. Li, G.F. Gao, Novel Immunodominant Peptide Presentation Strategy: a Featured HLA-A * 2402-Restricted Cytotoxic T-Lymphocyte Epitope Stabilized by Intrachain Hydrogen Bonds from Severe Acute Respiratory Syndrome Coronavirus Nucleocapsid Protein, J. Virol. 84 (2010) 11849–11857. https://doi.org/10.1128/JVI.01464-10.
- [46] N. Keicho, S. Itoyama, K. Kashiwase, N. Chi, H. Thuy, Association of human leukocyte antigen class II alleles with severe acute respiratory syndrome in the Vietnamese population, HIM. 70 (2009) 527–531. https://doi.org/10.1016/j.humimm.2009.05.006.

- Epidemiological and Genetic Correlates of Severe Acute Respiratory Syndrome Coronavirus Infection in the Hospital with the Highest Nosocomial Infection Rate in Taiwan in 2003, J. Clin. Microbiol. 44 (2006) 359–365. https://doi.org/10.1128/JCM.44.2.359.
- [48] S.S. Infection, S. Wang, K. Chen, M. Chen, W. Li, Y. Chen, Human-Leukocyte Antigen Class I Cw 1502 and Class II DR 0301 Genotypes Are Associated with Resistance to Severe Acute Respiratory, Viral Immunol. 24 (2011) 421–426. https://doi.org/10.1089/vim.2011.0024.
- [49] X. Tu, W. Po, Y. Zhai, Functional polymorphisms of the *CL2 and MBL genes cumulatively increase susceptibility to severe acute respiratory syndrome corona virus infection, J. Infect. 71 (2015) 101–109. https://doi.org/10.1016/j.jinf.2015.03.006.
- [50] M. Kikkert, Innate Immune Evasion by Human Res, atory RNA Viruses, J. Innate Immun. 12 (2020) 4–20. https://doi.org/10.1159/000503030.
- [51] N. Vabret, G.J. Britton, C. Gruber, S. H., de, J. Kim, M. Kuksin, R. Levantovsky, L. Malle, A. Moreira, M.D. Park, *et al.* Immunology of COVID-19: Current State of the Science, Immunity. 52 (2020) 910–941. https://doi.org/10.1016/j.immuni.2020.05.002.
- [52] E. De Wit, N. Van Doremalen, D. Falzarano, V.J. Munster, SARS and MERS: Recent insights into emerging coronaviruses, Nat. Rev. Microbiol. 14 (2016) 523–534. https://doi.org/10.1038/prr.nicro.2016.81.
- [53] E.J. Snijder, Y. van der Meer, J. Zevenhoven-Dobbe, J.J.M. Onderwater, J. van der Meulen, H.K. Koerten, A.M. Mommaas, Ultrastructure and Origin of Membrane Vesicles Associated with the Severe Acute Respiratory Syndrome Coronavirus Replication Complex, J. Virol. 80 (2006) 5927–5940. https://doi.org/10.1128/jvi.02501-05.
- [54] M.G. Wathelet, M. Orr, M.B. Frieman, R.S. Baric, Severe Acute Respiratory Syndrome Coronavirus Evades Antiviral Signaling: Role of nsp1 and Rational Design of an Attenuated Strain, J. Virol. 81 (2007) 11620–11633. https://doi.org/10.1128/jvi.00702-07.

- Drosten, M.A. Muller, Middle East Respiratory Syndrome Coronavirus Accessory Protein 4a Is a Type I Interferon Antagonist, J. Virol. 87 (2013) 12489–12495. https://doi.org/10.1128/jvi.01845-13.
- [56] Y. Yang, L. Zhang, H. Geng, Y. Deng, B. Huang, Y. Guo, Z. Zhao, W. Tan, The structural and accessory proteins M, ORF 4a, ORF 4b, and ORF 5 of Middle East respiratory syndrome coronavirus (MERS-CoV) are potent interferon antagonists, Protein Cell. 4 (2013) 951–961. https://doi.org/10.1007/s13238-013-3096-8.
- [57] S. Dong, J. Sun, Z. Mao, L. Wang, Y.L. Lu, J. Li, A guideline for homology modeling of the proteins from newly discovered betacoronavirus, 2019 novel coronavirus (. 019-nCoV), J. Med. Virol. (2020). https://doi.org/10.1002/jmv.25768.
- [58] F.C. Zhu, Y.H. Li, X.H. Guan, L.H. Hou, W.J. Wang, T.X. Li, S.P. Wu, B. Sen Wang, Z. Wang, L. Wang, *et al.* Safety, tolerability, and immunogenicity of a recombinant adenovirus type-5 vectored COVID-19 vaccine: a dose-escalation, open-label, non-randomised, first-in-human trial, Lancet. (2020). https://doi.org/10.1016/S0140-<736(20)31208-3.
- [59] R. Zhang, Y. Li, T.J. Cowley, A.D. Steinbrenner, J.M. Phillips, B.L. Yount, R.S. Baric, S.R. Weiss, The nsp1, nsp13, and M Protein. Contribute to the Hepatotropism of Murine Coronavirus JHM.WU, J. Virol. 89 (2015) 3598–2529. https://doi.org/10.1128/jvi.03535-14.
- [60] M. Hackbart, X. Deng, S.C. Baker, Coronavirus endoribonuclease targets viral polyuridine sequences to evade activating host sensors, Proc. Natl. Acad. Sci. U. S. A. 117 (2020) 8094–8103. https://doi.org/10.1073/pnas.1921485117.
- [61] M.N.I. kim young, Jedrzejczak robert, Crystal structure of Nsp15 endoribonuclease NendoU from SARS-CoV-2, Protein Sci. Publ. Protein Soc. 29 (2019) 1596–1605.
- [62] C.K. Yuen, J.Y. Lam, W.M. Wong, L.F. Mak, X. Wang, H. Chu, J.P. Cai, D.Y. Jin, K.K.W. To, et al. SARS-CoV-2 nsp13, nsp14, nsp15 and orf6 function as potent interferon antagonists, Emerg. Microbes Infect. 9 (2020) 1418–1428. https://doi.org/10.1080/22221751.2020.1780953.

- Pathogenicity of MERS-CoV Proteins, Engineering. 5 (2019) 940–947. https://doi.org/10.1016/j.eng.2018.11.035.
- [64] Q. Liang, J. Li, M. Guo, X. Tian, C. Liu, X. Wang, X. Yang, P. Wu, Z. Xiao, Y. Qu, *et al.* Virus-host interactome and proteomic survey of PMBCs from COVID-19 patients reveal potential virulence factors influencing SARS-CoV-2 pathogenesis, BioRxiv. (2020). https://doi.org/10.1101/2020.03.31.019216.
- [65] W. Aouadi, A. Blanjoie, J. Vasseur, B. Canard, E. Decroly, crossm Binding of the Methyl Donor, 91 (2017) 1–18.
- [66] G. Sutton, E. Fry, L. Carter, S. Sainsbury, T. Walter, J. Yettleship, N. Berrow, R. Owens, R. Gilbert, A. Davidson, *et al.* The nsp9 Replicase Protein of SARS-Coronavirus, Structure and Functional Insights, Structure. 12 (2004) 341–353. https://dci.org/10.1016/j.str.2004.01.016.
- [67] K.J. Jang, S. Jeong, D.Y. Kang, N. Sp, Y.Y. Yang, D.E. Kim, A high ATP concentration enhances the cooperative translocation of the SANS coronavirus helicase nsP13 in the unwinding of duplex RNA, Sci. Rep. 10 (2020) 1–13. http://uoi.org/10.1038/s41598-020-61432-1.
- [68] S. Kang, M. Yang, Z. Hong, L. Zhang, Z. Huang, X. Chen, S. He, Z. Zhou, Z. Zhou, Q. Chen, *et al.* Crystal structure of SAR1-CoV-2 nucleocapsid protein RNA binding domain reveals potential unique drug targeting sites, Acta harm. Sin. B. (2020). https://doi.org/10.1016/j.apsb.2020.04.009.
- [69] C.-S. Shi, H.-Y. Qi, C. Boularan, N.-N. Huang, M. Abu-Asab, J.H. Shelhamer, J.H. Kehrl, SARS-Coronavirus Open Reading Frame-9b Suppresses Innate Immunity by Targeting Mitochondria and the MAVS/TRAF3/TRAF6 Signalosome, J. Immunol. 193 (2014) 3080–3089. https://doi.org/10.4049/jimmunol.1303196.
- [70] D.E. Gordon, G.M. Jang, M. Bouhaddou, J. Xu, K. Obernier, K.M. White, M.J. O'Meara, V. V. Rezelj, J.Z. Guo, D.L. Swaney, *et al.* A SARS-CoV-2 protein interaction map reveals targets for drug repurposing, Nature. 583 (2020). https://doi.org/10.1038/s41586-020-2286-9.

[71]

- important research questions, Cell Biosci. 10 (2020) 1–5. https://doi.org/10.1186/s13578-020-00404-4.
- [72] V.D. Menachery, H.D. Mitchell, A.S. Cockrell, L.E. Gralinski, B.L. Yount, R.L. Graham, E.T. McAnarney, M.G. Douglas, T. Scobey, A. Beall, *et al.* MERS-CoV accessory orfs play key role for infection and pathogenesis, MBio. 8 (2017) 1–14. https://doi.org/10.1128/mBio.00665-17.
- [73] I.Y. Chen, M. Moriyama, M.F. Chang, T. Ichinohe, Severe acute respiratory syndrome coronavirus viroporin 3a activates the NLRP3 inflammasome, Front. Microbiol. 10 (2019) 1–9. https://doi.org/10.3389/fmicb.2019.00050.
- [74] M. Guilliams, B.N. Lambrecht, H. Hammad, Division of labor between lung dendritic cells and macrophages in the defense against pulmonary infections, Mucosal Immunol. 6 (2013) 464–473. https://doi.org/10.1038/mi.2013.14.
- [75] S. Perlman, A.A. Dandekar, Immunopath genesis of coronavirus infections: Implications for SARS, Nat. Rev. Immunol. 5 (2005) 917–927 https://doi.org/10.1038/nri1732.
- [76] B. Diao, Z. Feng, C. Wang, H. Vang, L. Liu, C. Wang, R. Wang, Y. Liu, Y. Liu, G. Wang, et al., Human Kidney is a Target for Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Infection, MedR2 iv. 2 (2020). https://doi.org/10.1101/2020.03.04.20031120.
- [77] W. Wen, W. Su, H. Tang, W. Le, X. Zhang, Y. Zheng, X. Liu, L. Xie, J. Li, J. Ye, *et al.* Immune cell pro fi ling of COVID-19 patients in the recovery stage by single-cell sequencing, Cell Discov. 6 (2020) 31. https://doi.org/10.1038/s41421-020-0168-9.
- [78] E.Z. Ong, Y. Fu, Z. Chan, W.Y. Leong, A. Bertoletti, E.E. Ooi, E.Z. Ong, Y. Fu, Z. Chan, W.Y. Leong, et al. Brief Report A Dynamic Immune Response Shapes COVID-19 Progression II II Brief Report A Dynamic Immune Response Shapes COVID-19 Progression, Cell Host Microbe. 27 (2020) 879-882.e2. https://doi.org/10.1016/j.chom.2020.03.021.
- [79] M. Liao, Y. Liu, J. Yuan, Y. Wen, G. Xu, J. Zhao, L. Chen, J. Li, X. Wang, F. Wang, et al. The

- sequencing, MedRxiv. (2020) 2020.02.23.20026690. https://doi.org/10.1101/2020.02.23.20026690.
- [80] J. Zhao, J. Zhao, N. Van Rooijen, S. Perlman, Evasion by Stealth: Inefficient Immune Activation Underlies Poor T Cell Response and Severe Disease in SARS-CoV-Infected Mice, PLoS Pathog. 5 (2009) e1000636. https://doi.org/10.1371/journal.ppat.1000636.
- [81] C. Page, L. Goicochea, K. Matthews, Y. Zhang, P. Klover, M.J. Holtzman, L. Hennighausen, M. Frieman, Induction of Alternatively Activated Macrophages Enhances Pathogenesis during Severe Acute Respiratory Syndrome Coronavirus, J. Virol. 86 (2012) 13334–13349. https://doi.org/10.1128/JVI.01689-12.
- [82] Z. Feng, B. Diao, R. Wang, G. Wang, C. Wang, Y. Tan, C. Wang, Y. Liu, Y. Liu, Z. Yuan, *et al.* The Novel Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) Directly Decimates Human Spleens and Lymph Nodes Running tit et SARS-CoV-2 infects human spleens and lymph nodes, MedRxiv. 2 (2020) 1–18.
- [83] J.H. Wenjun W, Xiaoqing L, Sipei W, Puyi L, Liyan H, Yimin L, Linling C, Sibei C, Lingbo N, Yongping L, The definition and risk of Cytokine Release Syndrome-Like in 11 COVID-19-Infected Pneumonia critically ill patient. Disease Characteristics and Retrospective Analysis, MedRxiv. (2020). https://doi.org/10.1171/2020.02.26.20026989.
- [84] M. Zheng, Y. Gao, G. Wang, G. Song, S. Liu, D. Sun, Y. Xu, Z. Tian, Functional exhaustion of antiviral lymphocytes in COVID-19 patients, Cell. Mol. Immunol. 17 (2020) 533–535. https://doi.org/10.1038/s41423-020-0402-2.
- [85] C.-Y. Song, J. Xu, J.-Q. He, Y.-Q. Lu, COVID-19 early warning score: a multi-parameter screening tool to identify highly suspected patients, MedRxiv. (2020). https://doi.org/10.1101/2020.03.05.20031906.
- [86] L.E. Carlin, E.A. Hemann, Z.R. Zacharias, J.W. Heusel, K.L. Legge, Natural killer cell recruitment to the lung during influenza A virus infection is dependent on CXCR3, CCR5, and virus exposure dose,

- [87] M. Liao, Y. Liu, J. Yuan, Y. Wen, G. Xu, J. Zhao, L. Cheng, J. Li, X. Wang, F. Wang, et al. Single-cell landscape of bronchoalveolar immune cells in patients with COVID-19, Nat. Med. 26 (2020) 842–844. https://doi.org/10.1038/s41591-020-0901-9.
- [88] F. Wang, J. Nie, H. Wang, Q. Zhao, Y. Xiong, L. Deng, S. Song, Z. Ma, P. Mo, Y. Zhang, Characteristics of Peripheral Lymphocyte Subset Alteration in COVID-19 Pneumonia, J. Infect. Dis. 221 (2020) 1762–1769. https://doi.org/10.1093/infdis/jiaa150.
- [89] G. Li, X. Chen, A. Xu, Profile of specific antibodies to the SARJ associated coronavirus, N. Engl. J. Med. 349 (2003) 508–509. https://doi.org/10.1056/NEJM20030.7313490520.
- [90] D.L. Thomas, COVID 19 progression linked to B cell activation, medRxiv (2020).
- [91] Z. Xu, L. Shi, Y. Wang, J. Zhang, L. Huang, C. Zhang, S. Liu, P. Zhao, H. Liu, L. Zhu, et al. Pathological findings of COVID-19 associated with acute respiratory distress syndrome, Lancet Respir. Med. 8 (2020) 420–422. https://doi.org/10.1016/S2213-2600(20)30076-X.
- [92] H. Zheng, M. Zhang, C. Yang, N. Zhang, X. Wang, X. Yang, X. Dong, Y. Zheng, Elevated exhaustion levels and reduced functional diversity of T cells in peripheral blood may predict severe progression in COVID-19 patients, Cell. Mol. Immunol. 17 (2020) 541–543. https://doi.org/10.1038/c41-123-020-0401-3.
- [93] Y.Y. Fan, Z.T. Huang, L. Li, M.H. Wu, T. Yu, R.A. Koup, R.T. Bailer, C.Y. Wu, Characterization of SARS-CoV-specific memory T cells from recovered individuals 4 years after infection, Arch. Virol. 154 (2009) 1093–1099. https://doi.org/10.1007/s00705-009-0409-6.
- [94] Y. Zhou, B. Fu, X. Zheng, D. Wang, C. Zhao, Y. Qi, R. Sun, Z. Tian, *et al.* Pathogenic T-cells and inflammatory monocytes incite inflammatory storms in severe COVID-19 patients, Natl. Sci. Rev. 7 (2020) 998–1002. https://doi.org/10.1093/nsr/nwaa041.
- [95] M. Liao, Y. Liu, J. Yuan, Y. Wen, G. Xu, J. Zhao, L. Chen, J. Li, X. Wang, F. Wang, et al. The landscape of lung bronchoalveolar immune cells in COVID-19 revealed by single-cell RNA

- [96] B. Diao, C. Wang, Y. Tan, X. Chen, Y. Liu, L. Ning, L. Chen, M. Li, Y. Liu, G. Wang, et al. Reduction and Functional Exhaustion of T Cells in Patients With Coronavirus Disease 2019 (COVID-19), Front. Immunol. 11 (2020) 827. https://doi.org/10.3389/fimmu.2020.00827.
- [97] C.K. Wong, C.W.K. Lam, A.K.L. Wu, W.K. Ip, N.L.S. Lee, I.H.S. Chan, L.C.W. Lit, D.S.C. Hui, M.H.M. Chan, S.S.C. Chung, *et al.* Plasma inflammatory cytokines and chemokines in severe acute respiratory syndrome, Clin. Exp. Immunol. 136 (2004) 95–103. https://doi.org/10.1111/j.1365-2249.2004.02415.x.
- [98] R. Channappanavar, A.R. Fehr, R. Vijay, M. Mack, J. Zhao, D.K. Meyerholz, S. Perlman, Dysregulated Type I Interferon and Inflammatory Monocyte-Macrophage Responses Cause Lethal Pneumonia in SARS-CoV-Infected Mice, Cen Host Microbe. 19 (2016) 181–193. https://doi.org/10.1016/j.chom.2016.01.007.
- [99] Y. Jiang, J. Xu, C. Zhou, Z. Wu, S. Zhong, J. Liu, W. Luo, T. Chen, Q. Qin, P. Deng, Characterization of cytokine/chemokine profiles of seven acute respiratory syndrome, Am. J. Respir. Crit. Care Med. 171 (2005) 850–857. https://doi.org/10.1164/rccm.200407-857OC.
- [100] F. Coperchini, L. Chiovato, L. Croce, F. Magri, M. Rotondi, The cytokine storm in COVID-19: An overview of the involven ent of the chemokine/chemokine-receptor system, Cytokine Growth Factor Rev. 53 (2020) 25–32. https://doi.org/10.1016/j.cytogfr.2020.05.003.
- [101] X. Li, M. Geng, Y. Peng, L. Meng, S. Lu, Molecular immune pathogenesis and diagnosis of COVID-19, J. Pharm. Anal. 10 (2020) 102–108. https://doi.org/10.1016/j.jpha.2020.03.001.
- [102] J.M. Sanders, M.L. Monogue, T.Z. Jodlowski, J.B. Cutrell, Pharmacologic Treatments for Coronavirus Disease 2019 (COVID-19) A Review, JAMA. 323 (2020) 1824–1836. https://doi.org/10.1001/jama.2020.6019.
- [103] C.A. Devaux, J.-M. Rolain, P. Colson, D. Raoult, New insights on the antiviral effects of chloroquine against coronavirus: what to expect for COVID-19?, Int. J. Antimicrob. Agents. (2020) 105938.

- [104] P. Colson, J.M. Rolain, J.C. Lagier, P. Brouqui, D. Raoult, Chloroquine and hydroxychloroquine as available weapons to fight COVID-19, Int. J. Antimicrob. Agents. 55 (2020) 105932. https://doi.org/10.1016/j.ijantimicag.2020.105932.
- [105] D. Zhou, S.-M. Dai, Q. Tong, COVID-19: a recommendation to examine the effect of hydroxychloroquine in preventing infection and progression, J. Antimicrob. Chemother. 75 (2020) 1667–1670. https://doi.org/10.1093/jac/dkaa114.
- [106] B. Cao, Y. Wang, D. Wen, W. Liu, J. Wang, G. Fan, L. Ruan, B. Cong, Y. Cai, M. Wei, *et al.* A Trial of Lopinavir-Ritonavir in Adults Hospitalized with Severe Cocid-19, N. Engl. J. Med. 382 (2020) 1787–1799. https://doi.org/10.1056/NEJMoa2001282.
- [107] J.A. Al-Tawfiq, A.H. Al-Homoud, Z.A. Memish, Reindesivir as a possible therapeutic option for the COVID-19, Travel Med. Infect. Dis. 34 (2020) 101515. https://doi.org/10.1016/j.tmaid.2020.101615.
- [108] R.U. Kadam, I.A. Wilson, Structural basis of influenza virus fusion inhibition by the antiviral drug Arbidol, Proc. Natl. Aca i. Sci. U. S. A. 114 (2017) 206–214. https://doi.org/10.1073/pnas.1617 \(\) 20114.
- [109] A. Casadevall, L. Pirofski, The convalescent sera option for containing COVID-19, J. Clin. Invest. 130 (2020) 1545–1548. https://doi.org/10.1172/jci138003.
- [110] M. Yuan, N.C. Wu, X. Zhu, C.C.D. Lee, R.T.Y. So, H. Lv, C.K.P. Mok, I.A. Wilson, A highly conserved cryptic epitope in the receptor binding domains of SARS-CoV-2 and SARS-CoV, Science (80-.). 368 (2020) 630–633. https://doi.org/10.1126/science.abb7269.
- [111] V. Baruah, S. Bose, Immunoinformatics-aided identification of T cell and B cell epitopes in the surface glycoprotein of 2019-nCoV, J. Med. Virol. 92 (2020) 495–500. https://doi.org/10.1002/jmv.25698.
- [112] J.R. Lon, Y. Bai, B. Zhong, F. Cai, H. Du, Prediction and Evolution of B Cell Epitopes of Surface Protein in SARS-CoV-2, BioRxiv. (2020). https://doi.org/10.1101/2020.04.03.022723.

- Discov. 5 (2020) 100019. https://doi.org/10.1016/j.medidd.2020.100019.
- [114] ALBERTA HOPE COVID-19 for the Prevention of Severe COVID19 Disease Full Text View -ClinicalTrials.gov, (n.d.).
- [115] Study to Evaluate the Safety and Antiviral Activity of Remdesivir (GS-5734TM) in Participants With Severe Coronavirus Disease (COVID-19) - Full Text View - ClinicalTrials.gov, (n.d.).
- [116] J.D. Goldman, D.C.B. Lye, D.S. Hui, K.M. Marks, R. Bruno, R. Montejano, C.D. Spinner, M. Galli, M.-Y. Ahn, R.G. Nahass, et al. Remdesivir for 5 or 10 Days in Patients with Severe Covid-19, N. Engl. J. Med. (2020). https://doi.org/10.1056/nejmoa2015301
- [117] C.D. Spinner, R.L. Gottlieb, G.J. Criner, J.R. Arribas L´vocz, A.M. Cattelan, A. Soriano Viladomiu, O. Ogbuagu, P. Malhotra, K.M. Mullane, A. Castag a, e, al. Effect of Remdesivir vs Standard Care on Clinical Status at 11 Days in Patients With Moderate COVID-19, JAMA. (2020). https://doi.org/10.1001/jama.2020.16349.
- [118] Duvelisib to Combat COVID-19 Full Tixt View Clinical Trials.gov, (n.d.).
- [119] Application of Desferal to Trea CO ID-19 Full Text View Clinical Trials.gov, (n.d.).
- [120] H. Wang, Z. Li, J. Niu, Y. Yu, L. Ma, A. Lu, X. Wang, Z. Qian, Z. Huang, X. Jin, et al. Antiviral effects of ferric ammonium citrate, Cell Discov. 4 (2018) 14. https://doi.org/10.1038/s41421-018-0013-6.
- [121] Clinical Study To Evaluate The Performance And Safety Of Favipiravir in COVID-19 Full Text View - ClinicalTrials.gov, (n.d.).
- [122] Study to Evaluate the Efficacy and Safety of Tocilizumab Versus Corticosteroids in Hospitalised COVID-19 Patients With High Risk of Progression - Full Text View - ClinicalTrials.gov, (n.d.).
- [123] Sarilumab COVID-19 Full Text View ClinicalTrials.gov, (n.d.).
- [124] J.J.V. McMurray, S.D. Solomon, S.E. Inzucchi, L. Kober, M.N. Kosiborod, F.A. Martinez, P.

- failure and reduced ejection fraction, N. Engl. J. Med. 381 (2019) 1995–2008. https://doi.org/10.1056/NEJMoa1911303.
- [125] Dapagliflozin in Respiratory Failure in Patients With COVID-19 Full Text View ClinicalTrials.gov, (n.d.).
- [126] Recombinant Human Angiotensin-converting Enzyme 2 (rhACE2) as a Treatment for Patients With COVID-19 Full Text View ClinicalTrials.gov, (n.d.).
- [127] The Phase 2 Study to Evaluate the Safety and Efficacy of Cavudine in Patients With Moderate COVID-19 Full Text View ClinicalTrials.gov, (n.d.).
- [128] Study of FT516 for the Treatment of COVID-19 in *Hamilian* Patients With Hypoxia Full Text View ClinicalTrials.gov, (n.d.).
- [129] DAS181 for Severe COVID-19: Compassion te Vise Full Text View Clinical Trials.gov, (n.d.).
- [130] Study of Open Label Losartan in COV₁?-19 Full Text View ClinicalTrials.gov, (n.d.).
- [131] C. Lv, W. Liu, B. Wang, R. Darg, Y. Qiu, J. Ren, C. Yan, Z. Yang, X. Wang, Ivermectin inhibits DNA polymerase UL42 of psquarabies virus entrance into the nucleus and proliferation of the virus in vitro and vivo, Antivira S. 159 (2018) 55–62. https://doi.org/10.1016/j.antiviral.2018.09.010.
- [132] S.K. Hong, H.J. Kim, C.S. Song, I.S. Choi, J.B. Lee, S.Y. Park, Nitazoxanide suppresses IL-6 production in LPS-stimulated mouse macrophages and TG-injected mice, Int. Immunopharmacol. 13 (2012) 23–27. https://doi.org/10.1016/j.intimp.2012.03.002.
- [133] Ivermectin and Nitazoxanide Combination Therapy for COVID-19 Full Text View ClinicalTrials.gov, (n.d.).
- [134] C. Shen, Z. Wang, F. Zhao, Y. Yang, J. Li, J. Yuan, F. Wang, D. Li, M. Yang, L. Xing, et al. Treatment of 5 Critically Ill Patients with COVID-19 with Convalescent Plasma, JAMA J. Am. Med. Assoc. 323 (2020) 1582–1589. https://doi.org/10.1001/jama.2020.4783.

- ClinicalTrials.gov, (n.d.).
- [136] Isotretinoin in Treatment of COVID-19 Full Text View Clinical Trials.gov, (n.d.).
- [137] Colchicine in COVID-19: a Pilot Study Full Text View Clinical Trials.gov, (n.d.).
- [138] Ruxolitinib in Covid-19 Patients With Defined Hyperinflammation Full Text View ClinicalTrials.gov, (n.d.).
- [139] Expanded Access Program of Ruxolitinib for the Emergency Treatment of Cytokine Storm From COVID-19 Infection Full Text View ClinicalTrials.gov, (n. i.).
- [140] F. La Rosée, H.C. Bremer, I. Gehrke, A. Kehr, A. Hochl aug. 5. Birndt, M. Fellhauer, M. Henkes, B. Kumle, S.G. Russo, P. La Rosée, The Janus kinase 1/2 inhibitor ruxolitinib in COVID-19 with severe systemic hyperinflammation, Leukemia. 34 (20%) 1805–1815. https://doi.org/10.1038/s41375-020-0891-0.
- [141] A Pilot Study of Sildenafil in COVID-1? Full Text View ClinicalTrials.gov, (n.d.).
- [142] Sirolimus Treatment in Hospita'ıze Patients With COVID-19 Pneumonia Full Text View ClinicalTrials.gov, (n.d.).
- [143] Single-Blind Study of a Single Dose of Peginterferon Lambda-1a Compared With Placebo in Outpatients With Mild COVID-19 Full Text View ClinicalTrials.gov, (n.d.).
- [144] Rintatolimod and IFN Alpha-2b for the Treatment of Mild or Moderate COVID-19 Infection in Cancer Patients Full Text View ClinicalTrials.gov, (n.d.).
- [145] Administration of Intravenous Vitamin C in Novel Coronavirus Infection (COVID-19) and Decreased Oxygenation Full Text View ClinicalTrials.gov, (n.d.).
- [146] Safety and Immunogenicity Study of 2019-nCoV Vaccine (mRNA-1273) for Prophylaxis of SARS-CoV-2 Infection (COVID-19) Full Text View ClinicalTrials.gov, (n.d.).
- [147] L.A. Jackson, E.J. Anderson, N.G. Rouphael, P.C. Roberts, M. Makhene, R.N. Coler, M.P.

- CoV-2 Preliminary Report, N. Engl. J. Med. (2020). https://doi.org/10.1056/nejmoa2022483.
- [148] Safety, Tolerability and Immunogenicity of INO-4800 for COVID-19 in Healthy Volunteers Full Text View ClinicalTrials.gov, (n.d.).
- [149] P.M. Folegatti, K.J. Ewer, P.K. Aley, B. Angus, S. Becker, S. Belij-Rammerstorfer, D. Bellamy, S. Bibi, M. Bittaye, E.A. Clutterbuck, *et al.* Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial, Lancet. 396 (2020) 467–478. https://doi.org/10.016/S0140-6736(20)31604-4.
- [150] A Study of a Candidate COVID-19 Vaccine (COV001) Full Vera View Clinical Trials.gov, (n.d.).
- [151] Immunity and Safety of Covid-19 Synthetic Minigene Voccine Full Text View ClinicalTrials.gov, (n.d.).
- [152] Evaluation of the Safety and Immunography of a SARS-CoV-2 rS (COVID-19) Nanoparticle Vaccine With/Without Matrix-M Adjuvant Full Text View ClinicalTrials.gov, (n.d.).
- [153] Study to Describe the Safety, Tolerao (ity, Immunogenicity, and Potential Efficacy of RNA Vaccine Candidates Against COVID-19 n healthy Adults Full Text View ClinicalTrials.gov, (n.d.).
- [154] Evaluating the Safety, Teler, buity and Immunogenicity of bacTRL-Spike Vaccine for Prevention of COVID-19 Full Text View ClinicalTrials.gov, (n.d.).

Journal	Pre-proof
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ble 1: Vario

utations	Features	Outcomes	Ref.
ikeD614G	A missense mutation in S protein encoding gene, where an amino acid (aa) change from aspartate to glycine at 614 position was found. With this mutation this strain contains 3 other mutations as follow: 1. C-to-T mutation at the 5' untranslated region (UTR) at position 241, 2. C-to-T mutation at position 3037, 3. a nonsynonymous C-to T mutation at positior 1-1408 within the RNA dependent RNA polymera, e gene.	D614G substitution was a rare mutation at the beginning of the COVID-19 spread before March 2020, found as predominant in Europe, but later it occurred about 74% in all published sequences in June 2020 and spread worldwide. This production enhances the viral replication in the upper respiratory tract and and has higher susceptible to neut 31/12 tion by monoclonal antibodies.	[16,17]
SP2 and SP3	SARS-CoV-2 contains a polar aa instead of nonpolar aa unlike bat SARS at position 321 and glycine is replaced by serine in NSP3 at position of 543.	This may affect the mechanisms involved in viral entry and replication and increases the contagiousness of the virus.	[18]

RS-CoV-2	Journ	nal Pre-proof	
2012/01	CoV-2 has 29 aa substitution from the original Wuhan strain with a mutation N501Y which is located in the receptor binding region.	According to till now revealed reports this new strain possesses a high transmissible rate than the original strain.	
utations Rdrp	A mutation was found in the RNA dependent RNA polymerase at the position of 14408.	It might result in drug resistant viral phenotype.	[20]
utations ORF gion	According to the present studies there are mutations in ORF region as follow: 1. C > T in ORF1ab gene at position 8782, 2. T > C in ORF8 gene at position 28144.	Better studies needed to unc'erstand the role of this nutation in viral pathogenecity.	[21]
82 riant	This variant has 382 nucleotide deletion in ORF8.	This variant, seen during the early epidemic in Wuhan, was mild infectious with lower concentration of proinflammatory cytokines.	[22]

RS-CoV-	Journal Pre-proof					
AZ-	of 81 nucleotide in the	to be studied.				
SU2923	ORF7a region found in					
	Arizona.					



ame of the	Structure	ructure Function on immunological aspects						
ructural otein		SARS-CoV-1	MERS	SARS-CoV-2				
nstructural	It is a leader protein,	Antagonize IFN-β	Helps in viral	Detailed function				
otein (nsp)1 -59]	cleaved chain of ORF1b.	production by decreasing the	replication.	not known.				
67]		phosphorylation level of STAT1.	Ö					
sp 15 [60–64]	Nidoviral RNA	Cleaves polyuridine	Provent	Inhibits the				
	uridylate specific	(polyU) sequences from	civation of	nuclear				
	endoribonuclease	PUN RNAs and hants	dsRNA sensors	localization of				
	(NendoU) that	the formation of a	in host cell for	IFR3 and				
	belongs to EndoU family.	PAMP and thus impedes the ability of	evading immune system.	antagonize the production of IFN				
		ac valion the innate	System.	and also target				
	34 KDa, around 345 amino acids, wit'.	mr une response to		RNF41 (also				
	three domains: N	infection by MDA5.		known as NRDP1)				
	terminal, middle and			to regulate innate				
	C-terminal domain.			immune system.				
sp 9 and Nsp	The crystalline	Nsp10 regulates the	According to	Interacts with NF-				
[64–66]	structure of Nsp9 of	activity of the 2'-O-	available data	κB repressor,				
	SARS-CoV revealed	Methyltransferase (2'-	Nsp9 helps in	NKRF and				
	that the molecule form two distinct types of	O- MTase) that prevents virus detection	viral replication.	activates IL-8/IL-6 mediated				

		Journal Pre-pi	roof	
	of the protein is an	mechanisms and viral		neutrophils that
	open 6-stranded β-	translation inhibition by		results in
	barrel that in turn	the interferon-		inflammatory
	comprises of two	stimulated IFIT-1		response in
	antiparallel β sheets	protein.		patients.
	packed orthogon.			
p13	It is a helicase of	It acts as a helicase and	Nsp13 attenuates	Targeted TBK1
3,64][67]	superfamily 1 and	helps in unfold the	the viral	and TBKBP1 to
	helps in viral RNA	RNA-DNA hybrid.	rep icai on.	inhibit interferon
	replication via			pathway to
	unwinding of duplex		8	regulate innate
	RNA and DNA			immune response
	leaving a 5' single-			in host cell.
	stranded tail in a 5' to			
	3' direction.			
(Nucleocapsid)	N protein of SARS-	Generation of IFN is	Interacts with	Detail function is
otein [68]	CoV-2 is 29.9 kb n	retarded upon crosstalk	TRIM25 and	not clearly known.
	length, similarly tr	between the SPRY	interfere the IFN	
	27.9 kb SARS-CoV	domain of TRIM25 and	production in	
	and 30.1 kb MERS-	C terminus of the N	host cell.	
	CoV genome.	protein as it blocked		
		RIG-I ubiquitination by		
		TRIM25.		
RF (open	ORF-9b possess a	ORF-9b manipulates	Function in	ORF9b in
ading frame)	long hydrophobic	host cell mitochondrial	immune system	association with

[69,70]		Journal Pre-p	roof	
	formed due to interwined dimer with an amphipathic outer surface	MAVS signaling that results in reducing NLRP3 inflammasome activity, thus evading innate immune system.		with a signaling adaptor MAVS indirectly
RF6 [62,63]	SARS-CoV-1 and SARS-CoV-2 share only 69% amino acid similarity	It prevents primary production of interferon.	Helps in viral assembly and viral release and car art as a potential B cell epitope.	It prevents interferon production by various signaling molecules MDA5, MAVS, TBK1 and IRF3-5D, which is a phospho-mimic of the activated form of IRF3.
RF3 [71–73]	Accessory protein formed by the cleavage of ORF1 and ORF1b.	OPF3a is responsible For activation of the NLRP3 inflammasome by secreting IL-1β.	Prevent interferon production and prevent inflammation.	The hypothesis is that ORF3a of SARS-CoV-2 may be less efficient in inflammasome activation.

IFIT, interferon-induced protein with tetratricopeptide repeats; IFN, interferon; IRF, interferon regulatory factor; MTase, methyltransferase; N, nucleocapsid; NendoU, nidoviral RNA uridylate specific endoribonuclease; NF-κB, nuclear factor **kappa-**light-chain-enhancer of activated B cells; NLRP3, NLR family pyrin domain containing 3; NRDP1, neuregulin receptor degradation protein 1; Nsps, non-structural proteins; ORFs, open reading frames; PAMP, pathogen associated molecular patterns; PolyU,

transducer and activator of transcription; TBK1, TANK binding kinase 1; TBKBP1, TANK binding kinase-

1, binding protein 1; TRIM25, tripartite motif containing 25

	Function		
These drugs are basically used	Chloroquine and hydroxychloroquine		
in malaria treatment and some extent to Systemic Lupus	inhibits viral entry into cells. The		
Erythematosous (SLE) and	glycosylation of host receptors,		
rheumatoid arthritis (RA)	endosomal		
treatment.	acidification and proteolytic processing		
	are inhibited. These agents also affect		
	ıı. munopathology		
	via production of cytokine inhibition of		
	lysosomal activity and		
	autophagy of immune cells.		
Lopinavir and ritonavir are	No published data are available but		
approved by US Food and Drug	invitro studies show that they act by		
Administration (FDA) and in	inhibiting 3-chymotrypsin-		
treatm ent of HIV.	like protease.		
Remdesivir, also called GS-	The drug was designed against		
5734, is a monophosphate	microbes with activity also against		
prodrug that forms an active C-	RNA viruses. Remdesivir targets the		
adenosine nucleoside	RNA dependent RNA polymerase and		
triphosphate analogue	hamper the replication cycle of RNA		
undergoing metabolism.	viruses. Remdesivir first used for the		
	in malaria treatment and some extent to Systemic Lupus Erythematosous (SLE) and rheumatoid arthritis (RA) treatment. Lopinavir and ritonavir are approved by US Food and Drug Administration (FDA) and in treatment of HIV. Remdesivir, also called GS-5734, is a monophosphate prodrug that forms an active C-adenosine nucleoside triphosphate analogue		

	Jou	-pi	roof	
Umifenovir [108]	Umifenovir or	Arbid, a	ın	It inhibits S protein/ACE2 interaction
Chinenovii [100]	Chinenovii oi	Aibid, a	111	n minores 5 protein/ACL2 interaction
	antiviral drug.			via blocking the fusion of membrane
				with the viral envelope. Arbid is used
				for the treatment of influenza in Russia
				and China and is recently in the interest
				for treat.ng COVID-19.

ACE2, angiotensin converting enzyme 2; HIV, human immunodeficionary virus; RA, rheumatoid arthritis; S, spike; SLE, systemic lupus erythematosus; FDA, food and drug dministration

Candidate	Mode of action and	Existing	Trial	Location	Expected	Phase
drug	dose	disease approval	sponsor		Result	trial
		арргочаг				
Hydroxycholo	Hydroxychloroquine	Malaria,	Dr.	University	Preliminary trials	Phase
roquine	inhibits acidification of	rheumatoid	Michael	of Calgary	indicated that it is a	III
(ID:	endosomes,	arthritis	Hill		potential and safe drug	
NCT04329611	deglycosylates	(RA), lupus		4	against COVID-19	
)[114]	receptors of recipient			0	pneumonia and shorten	
<i>)</i> [114]	cells, prevents		4	\bigcirc	the disease course about	
	proteolytic processing		-0		50%. Later it was found	
	thus retards entry of		(7)		that both these drugs	
	virus. Inhibition of				have side effects like	
	cytokine production				allergic reactions,	
	modulates the host	0			hypoglycemia,	
	immune system. It also				cardiomyopathy. On	
	inhibits in host				April, 2020 in Brazil 11	
	autophagy				patients died due to	
	lysosomal functionality				irregular heart rates.	
	Dose:					
	Hydroxychloroquine					
	dose of 400 mg po bid					
	on day 1 followed by					

		Journal	l Pre-proo	f		
	be given twice daily for 4 days					
Remdesivir (ID:NCT0429 2899 and NCT04292730) [115–117]	Dose: RDV 200 mg on first Day followed by RDV 100 mg for next 4 days together with standard therapy.		Gilead, WHO, INSERM	China, japan	According to US NIAID, remdesivir shows faster recover from COVID-19 in 11 days compared to other drugs. A clinical trial in china, reported on 29 th April several adverse effect of remdesivir in treated patients.	In April 2020, there was 9 phase III clinical trials across the world.
Duvelisib (ID:NCT0437 2602) [118]	Target PI3K and control hyperactivation of innate immune system by affecting macrophage polarization, reducing inflammation in pulmonary and limit the persistence of viral load.		Washingto n University School of Medicine.	Washingto n University School of Medicine, Saint Luis, Missouri, United Sates	outcome reported on 30 th April overall	Phase

		Journa	l Pre-proo	f		
	for 10days, orally					
Deferoxamine	It is a natural product		Kermansh	Regenerati	Trial ongoing	Phase I
(ID:	which is isolated from		ah	ve		Phase
NCT04333550	Streptomyces pilosus. It		University	Medicine		II
) [119,120]	helps in the formation		of	Research		
	of iron complexes and		Medical	Center,		
	its mesylate form		Science	Kermansh		
	perform as chelating			ah		
	agent,			Um /ersity		
				υf		
				Medical		
				Science,		
				Iran,		
				kemansha		
		.00		h.		
Farininaria	It towarts DNIA	Tood hofom	Giuliano	Asst	the normalization of	Phase
Favipiravir	It targets RNA-	sed before				
(ID:NCT0433	dependent KNA	against	Rizzardini	Fatebenefr	pyrexia, normal	III
6904)[121]	polymerase (R-1, p)	Ebola virus		atelli	respiratory rate and	
	enzymes, which are	and lassa		Sacco,	relief from cough is	
	necessary for the	virus.		Milan	maintained for at least	
	transcription and			Ilaty	72 hours.	
	replication of viral					
	genomes.					
	Dose: Day 1:1800mg,					
	genomes.					

		Journa	l Pre-proo	f		
	on day 1 and day 2 followed by TID dose					
	at 600mg for 14 days.					
	Thereafter: 600mg,					
	TID, for a maximum of					
	14 days.					
Tocilizumab	Human	This drug	Genentech	Multiple	As per 8-point WHO	Phase
(NCT0434544	monoclonal antibody	has been	-hoffmann	cormiies	scale, improvement of	II
5) [122]	against IL-6 receptor.	used against	La Roche	O	more than 2 point is	
		immune			observed.	
		suppression				
	Dose: Intravenously	and in RA.				
	administered with a					
	concentration of 8					
	mg/kg (body weight)					
	once, within 60					
	minutes.					
Sarilumab	Human monoc¹c ıal	RA	Regeneron	Multiple	Patients improvement in	Phase
(ID:NCT0432	antibody against IL-6		-Sanofi	countries	oxygenation.	II/
7388) [123]	receptor.					Phase
, 6 6 6) [126]						III
	Dose: Sarilumab Dose					
	1 given intravenously					
	one time on Day 1					

Dapagliflo		Journa	l Pre-proo	f	1	Phase
(ID:NCT0435	glucose cotransporter	mia	Luke's	countries	functionality of organs	III
0593)	inhibitor.		Mid		are observed in	
[124,125]			America		hospitalized patients at	
[124,123]			Heart		30 th day.	
	Dose: Dapagliflozin 10		Institute,			
	mg daily		Astrazene			
			ca			
			Cu	×		
Recombinant	It is a monocarboxy-		Hospital	Gi ang lon	24-48 hours of	Phase
Human	peptidase that		of	¿ China	Pulmonary imaging	II
Angiotensin-	metabolizes several		Guang ho		showed that progression	
converting	peptides, including the		u w dical		of the lesions are more	
Enzyme 2	degradation of		University		than 50% and the	
(rhACE2)	angiotensin II, and				patients were managed	
(ID:NCT0428	contributes to	0			as severe	
7686)[126]	cardiovascular effect.					
	Danie Taradan M					
	Dose: Together with					
	standard treatment0.4					
	mg/kg IV BID given for					
	7 days.					
Clevudine	Clevudine is an	Hepatitis B	Bukwang		Trial ongoing	Phase
with	antiviral drug used		Pharmace			II
combination	against hepatitis B.		utical			
Hydroxychlor						

oquine		Journa	l Pre-proo	f		
(ID:NCT0434 7915) [127]	Dose: Clevudine 120 mg once daily for 14 days (Hydroxychloroquine 200mg twice daily for 14 days.					
Drug: FT516 (ID: NCT04363346) [128]	FT516 is a cryopreserved NK cell product of an iPSC that was transduced with ADAM17 non-cleavable CD16 (Fc receptor). Dose: Firstly, FT516 is administered at 9 × 1.07 cells/dose in low concentration Secondly, FT516 is first given at low dose (9 x107 cells/dose) additionally at Day 4 it is provided in medium	Cancer	Masonic Cancer Center, Univer ity of Ninnesota	Mir. reapol is, Mir.nesota , United States.	Trial ongoing	Phase I

cells/dose) Thirdly, along with the low and medium doses, a higher dose of drug is given at day 7 (9 x 108 cells/dose) Renmin Wu. 2n, Trial ongoing	
low and medium doses, a higher dose of drug is given at day 7 (9 x 108 cells/dose) Renmin Wu.van, Trial ongoing	
a higher dose of drug is given at day 7 (9 x 108 cells/dose) Renmin Wu. 2n, Trial ongoing	
given at day 7 (9 x 108 cells/dose) Renmin Wu. 2n, Trial ongoing	
Cells/dose) DAS181 (ID: Renmin Wu. 9n, Trial ongoing	
DAS181 (ID: Renmin Wullen, Trial ongoing	
NCT04324489 Hospital Ht bei,	
of Wuhar China	
Dose: 9mg daily for 10 Univer ity	
days	
Losartan University Kansas Trial ongoing Pha	nase I
(ID: NCT043 of Kansas City,	
35123) [130] Medical United	
Dose: 25 mg for first 3 Center States.	
days followed by 50	
mg QD till study	
completion	
Ivermectin Antiviral drug that Tanta Trial ongoing Phase	nase
with affect the viral RNA University II	
Nitazoxanide and DNA replication in	
(ID:	
NCT04360356	
Dose: Ivermectin 200	
mcg/kg once orally on	

		Journa	l Pre-proo	f		
	Nitazoxanide 500 mg					
	twice daily orally with					
	meal for 6 days					
Transfusion			Direction	France	Trial ongoing	Phase
of SARS-	Convalescent plasma		Centrale			III
CoV-2	contains antibody		du Service			
Convalescent	against SARS-CoV-2.		de Santé	C.		
Plasma.			des			
(ID:			Armées	0		
NCT04372979	Dose:					
) [134,135]	Intravenous		.0)			
Isotretinoin	Inhibitors of PLpro, a	used to	Tanta		Clinical clearance	Phase
(ID:NCT0436	protein encoded by	decrease	University		Change in COVID-19	III
1422) [136]	SARS-CoV2	virema in			virus load	
	Dose: Orally	ratients				
Colchicine	Non-selective inhibition	Cardiovascu	University	Italy	Trial ongoing	Phase
(ID:NCT0437	of NLRP3, a	lar disease	of Perugia			II
5202) [137]	pathophysiologic					
/	component of SARS-					
	CoV					
	Dose: 0.5mg every 8					

		Journa	l Pre-proo	f		
	orally (Tablet)					
Ruxolitinib	Treat the cytokine	Treat bone	Incyte	USA	Reduce 25%	Phase
(ID:	storm and	marrow	corporatio		hyperinflammation	II
NCT04355793	hyperinflammation in	disorders	n		caused due to the	
,NCT0433895	COVID-19 patients	like	University		cytokine storm	
8) [138–140]		myelofibros	of Jena			
	Dose: 5mg orally twice daily	Is		Ö		
	dany		2	O		
Sildenafil	Relaxes the muscles of	Erectile	Tongji	China	Respiratory symptom	Phase
(ID:NCT0430	the lungs by increasing	dysfunction	hospital		remission	III
4313) [141]	the potency of nitric				Decrease in fever	
	oxide gas to widen the					
	blood vessels resulting				C-reactive protein	
	in more oxygen	.00			recovery	
	inhalation					
		J '				
	Dose: 0.1g daily for 14					
	days, orally					
Sirolimus (ID:	mTOR inhibitor,	Used for	University	USA	Trail ongoing	Phase
NCT04341675	in or innotes,	preventing	of	ODII	Than ongoing	II
) [142]	immune suppressor	organ	Cincinnati			
)[142]		transplant	Cilicinian			
	Dagaséma on first Day	rejection				
	Dose:6mg on first Day					
	then 2mg daily for next	una				

		Journa	l Pre-proo	f		
		eiomyomato				
		sis (LAM)				
Peginterferon	Reduces viral shedding	Hepatitis B	Stanford	USA	Trial ongoing	Phase
Lambda-1a	of SARS-CoV-2	Virus	University			II
(ID:		infection				
NCT04331899	Danie	Hepatitis C				
) [143]	Dose: One	virus		Q.		
	subcutaneous injection	infection				
	of 180 ug			O		
Rintatolimod	Rintatolimod is a	Viral	Roswel'.	JSA	Trial ongoing	Phase
and IFN	dsRNA designed to	infections	Park			I/ IIa
Alpha-	mimic viral infection by		Center			
2b(ID:NCT04	activating immune		Cancer			
379518) [144]	pathways and IFN		Institute			
	Alpha-2b activate					
	immune responses and					
	both participate in					
	limiting viral					
	replication and					
	shedding					
	Dose: IV rintatolimod					
	for 2.5-3 hours together					
	with IV of recombinant					
	interferon alpha-2b over					
	I	<u> </u>	<u> </u>	<u>I</u>	1	

		Journa	l Pre-proo	f		
	3, 5, and 8 if there will					
	be the disease					
	progression or no					
	unacceptable toxicity					
	treatment will be					
	followed up at 14 th day					
	and 28 th day					
L-ascorbic	Reduce inflammation,	Sepsis	Hunter	Virginia,	Trial ongoing	Phase
acid (ID:	ARDS, reduce	_	Holmes	US.1		I/ II
NCT04357782	supplement		Mcguire			
) [145]	oxygenation, reduce		Vetera 1			
	risk respiratory failure		Afrais			
	which intubation		Medical			
			Center			
	Dose: 50mg/kg IV					
	given every 6 hours fo.	5,				
	4 days (16 total acres),					
mRNA1273	A lipid nanoparticle		National		Trial ongoing	Phase I
(ID:NCT0428	(LNP) encapsulated		Institute			
3461)	with mRNA encoding		of Allergy			
[146,147]	full length S protein of		and			
	SARS-CoV-2.		Infectious			
			Disease			
	Dece:10/25/50/100/250		(NIAID)			
	Dose: 10/25/50/100/250					

		Journa	l Pre-proo	f		
	.05mL intramuscular					
	injection in deltoid					
	muscle.					
INO-4880	It is a DNA vaccine		Inovio		Trial ongoing	Phase I
(ID:NCT0433	against whole- length S		Pharmace			
6410) [148]	protein of SARS-CoV-		uticals			
, , ,	2.			C.		
			_	O		
	Dose: intradermal					
	injection of 1.0 mg of					
	INO-4800		(Q)			
ChAdOx1	Adenovirus encoding		University	UK	Trial ongoing	Phase
nCoV-19	full-length S protein		of Oxford			II
2001 25	and an engine a process					
COVID-19						
(ID:NCT0432	Dose: One dose of 5 x					
4606)	10^10vp					
[149,150]	3					
COVID-19	Lentivirus infected		Shenzhen		Trial ongoing	Phase
LV- SMENP-	dendritic cells with		Geno-			II
DC	SMENP minigenes that		Immune			
(ID:NCT0427	express COVID-19		Medical			
6896) [151]	antigens and activated		Institute			
	CTLs.					

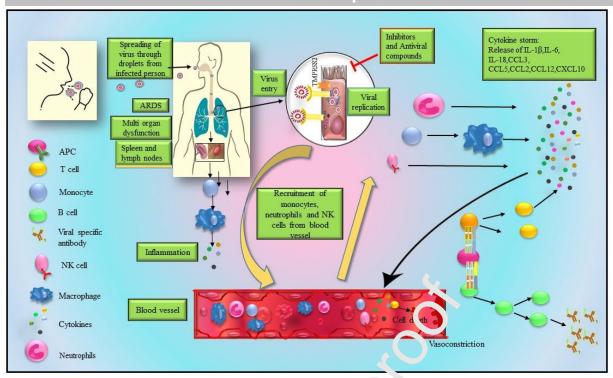
SARS-Cov		Journa	l Pre-proo	f		Phase I
(ID:NCT0436 8988) [152]	spike (S) protein of SARS-CoV-2					
BNT162a1,	It is a LNP		BioNTech		Trial ongoing	Phase I
b1, b2, c2	encapsulated		SE and			
(ID:NCT0436 8728)[153]	mRNA vaccines with mRNA targets for both larger S sequence and		Pfizer, Inc.	×.		
	RBD.					
	Dose: 0.5mL intramuscular injection.	Q				
Recombinant	Adenovirus type 5		CanSino	China	The vaccine is tolerable	Phase I
Novel	encoded with full	20	Biologics,		and immunogenic at 28	
Coronavirus	length S protein		Inc.		days post-vaccination in	
Vaccine		5			healthy adults, and rapid	
(Adenovirus	3				specific T-cell responses	
Type 5					were noted from day 14	
Vector)					post-vaccination.	
(ID:NCT0431						
3127) [58]						
bacTRL-	Live		Symvivo		Trail ongoing	Phase I
Spike-1	Bifidobacteriumlongum		Corporatio			
(ID:NCT0433	, engineered for the		n			

4980) [154		Journa	l Pre-proo	f	
	containing synthetic				
	DNA encoding spike				
	protein from SARS-				
	CoV-2.				

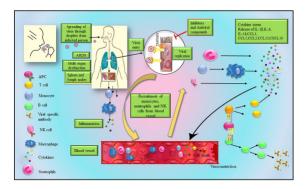
ADAM 17, a disintegrin and metalloproteinase 17; CTLs, cytotoxic T lymphocytes; dsRNA, double stranded RNA;IFN, interferon; LAM, lymphangioleiomyomatosis; LNP, lipid nanoparticle; mTOR, mammalian target of rapamycin; NK, natural killer; NLRP 3, NLR family pyrin domain containing 3; PI3K, phosphatidylinositol 3 kinase; RBD, receptor binding domain; RA, theumatoid arthritis; RdRp, RNA dependent RNA polymerase; S, spike;SARS-CoV-2, severe acute respiratory syndrome coronavirus 2



Journal Pre-proof



- Mutation accumulation results in new variants of SARS-CoV-2 causing severe pathogenicity.
- SARS-CoV-2 infection the apoptosis of lymphocytes via tissue resident CD169+ macrophages.
- In SARS-CoV-2 infection NK cells shows high level of negative immune check point marker.
- Epitope mapping and stem cell therapy are now also considered as a novel approach for drug development.
- Clinical trials of various repurposed drugs are ongoing for developing potent vaccines



Graphics Abstract

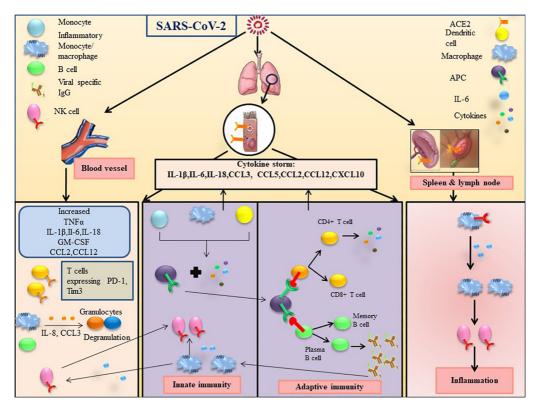


Figure 1

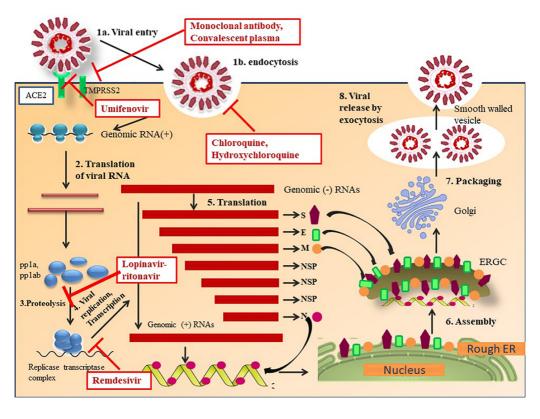


Figure 2